



Three-dimensional printing for craniomaxillofacial regeneration

Laura Gaviria, Joseph J. Pearson, Sergio A. Montelongo, Teja Guda, Joo L. Ong

Department of Biomedical Engineering, College of Engineering, The University of Texas at San Antonio, San Antonio, TX, USA

Abstract (J Korean Assoc Oral Maxillofac Surg 2017;43:288-298)

Craniomaxillofacial injuries produce complex wound environments involving various tissue types and treatment strategies. In a clinical setting, care is taken to properly irrigate and stabilize the injury, while grafts are molded in an attempt to maintain physiological functionality and cosmesis. This often requires multiple surgeries and grafts leading to added discomfort, pain and financial burden. Many of these injuries can lead to disfigurement and resultant loss of system function including mastication, respiration, and articulation, and these can lead to acute and long-term psychological impact on the patient. A main causality of these issues is the lack of an ability to spatially control pre-injury morphology while maintaining shape and function. With the advent of additive manufacturing (three-dimensional printing) and its use in conjunction with biomaterial regenerative strategies and stem cell research, there is an increased potential capacity to alleviate such limitations. This review focuses on the current capabilities of additive manufacturing platforms, completed research and potential for future uses in the treatment of craniomaxillofacial injuries, with an in-depth discussion of regeneration of the periodontal complex and teeth.

Key words: Three-dimensional printing, Periodontium, Hydroxyapatite, Biomaterials

[paper submitted 2017. 8. 31 / accepted 2017. 9. 11]

I. Introduction

Although the patterns of incidence and their causes have changed over the decades, craniomaxillofacial (CMF) injuries still occur worldwide. The most common causes include traffic and sports-related accidents, assaults, falls, civilian warfare¹⁻³, as well as diseases, congenital disorders and surgery⁴. CMF injuries are typically characterized by bone fractures in the frontal, orbital, nasal, maxillary and mandibular regions² and soft tissue damage such as complex lacerations, tissue avulsions, nerve and vessel injuries, and burns⁵⁻⁹. These complex injuries can compromise vital structures. Consequences of CMF injuries are disfigurement and dysfunction, including compromised airway, hemorrhaging, infection, scarring,

nerve damage, and non-union fractures². These disfigurement and dysfunction contribute to acute and long-term psychological problems as well to social and economic burdens^{8,10,11} since these detrimental outcomes are largely due to lack of full restoration of function and aesthetics found in the available treatments.

Due to the complexity of CMF injuries, the affected hard and soft tissues within the wound environment are unsuitable to support proper healing^{4,12,13}. Moreover, management of CMF injuries is extremely challenging and involves a multidisciplinary team of professionals for the treatment of facial bone fractures, dentoalveolar trauma, and soft tissue injuries as well as associated injuries, mainly to the head and neck regions^{3,11,14,15}. Major bone and soft tissue reconstruction often requires the use of autografts or allografts. Although autografts—considered the “gold standard”—and allografts are very attractive for their resorption, mechanical properties and immunological characteristics, both approaches have multiple limitations related to tissue availability, donor site morbidity and infection^{4,16-19}. Since major drawbacks for using autografts and allografts include the need to manually sculpture the grafts in the desired shape^{4,20}, synthetic alternatives using additive manufacturing have become an attractive option^{4,12,17}.

Joo L. Ong

Department of Biomedical Engineering, College of Engineering, The University of Texas at San Antonio, One UTSA Circle, San Antonio, TX 78249, USA

TEL: +1-210-458-7208 FAX: +1-210-458-5515

E-mail: anson.ong@utsa.edu

ORCID: <http://orcid.org/0000-0003-3330-2390>

© This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Copyright © 2017 The Korean Association of Oral and Maxillofacial Surgeons. All rights reserved.

As mentioned before, the primary goals of CMF repair are restoration of aesthetics and function, both requiring precise pre-surgical planning as well as prostheses and implants fabricated in very unique geometries and sizes^{15,18,20,21}. In the past decade, techniques such as additive manufacturing (e.g., three-dimensional [3D] printing) have been explored for tissue engineering purposes, especially for dental and CMF repair. This review focuses on tissue regenerative strategies for the CMF as a whole along with a focused discussion on 1) the regeneration of the periodontium and teeth within the oral cavity, and 2) providing an outlook on the advantages and limitations of current additive manufacturing, treatments and tissue regenerative research. The ability to harness the successes of tissue regeneration within specific regions of the CMF, such as the periodontium and teeth, could lead to a combined approach for regeneration of a larger region. This review will also discuss that potential and the ability of 3D printers to create a platform for manufacturing rather than the multiple manual manufacturing techniques currently used.

II. 3D Printing Technology for CMF Surgery

Additive manufacturing techniques, such as 3D printing, use the process of joining materials to create objects from digital 3D model data²⁰. For biomedical applications, 3D printing can be used for the fabrication of complex scaffold shapes that are specific to patients using computer aided design (CAD) and advanced medical imaging techniques such as magnetic resonance imaging and computed tomography (CT)^{12,18,19,22-29}.(Fig. 1)

Although many industries have benefited from the development of additive manufacturing technologies since the mid-1980s, their applications in the biomedical field have been slow due to technical challenges such as limited accuracy, low mechanical properties and lack of biomaterial

availability. All of these limitations have been investigated over the last two decades in order to address stringent performance and safety concerns^{12,19,22,27}. As a consequence, 3D printing technology has become more popular in tissue engineering and has found many applications in the fabrication of custom implants for the reconstruction of CMF defects²⁶. This has allowed for precise adaptation of the implant to the region of implantation, reducing surgical times and leading to lesser chances for infection, faster recovery and better cosmetics^{15,18,19,25,28}. 3D printing has also been introduced into the surgical field as a tool for pre-surgical planning^{25,29}, allowing surgeons to review and interact with the anatomical models, thereby facilitating the understanding of the morphology and making it easier to perform complex surgeries in less time^{18,24,27,30,31}. The uses of 3D printing for preoperative planning have been previously described in literature^{26,32}. While these planning techniques have expanded the knowledge in both the scientific and medical communities, the use of 3D printing towards tissue regeneration focuses on the need for specific biomaterial-based printing rather than rapid prototyping for surgical guidance.

1. 3D printed biomaterials for CMF repair

Early 3D printing research focused on the use of metals and ceramics¹² for bone tissue engineering. Ceramic scaffolds have been 3D printed and tested *in vitro* under static and dynamic conditions, achieving high printing resolution, structural mechanical support and cell growth^{20,23,33}. Today, 3D printing applications are investigated not only for bone reconstruction but also for replacement of soft tissues, using a variety of synthetic and biological materials including metals, ceramics and polymers^{12,18,20,22-24,27-29,33}. Although most known biomaterials can be processed using 3D printing, extensive optimization of processing and post processing

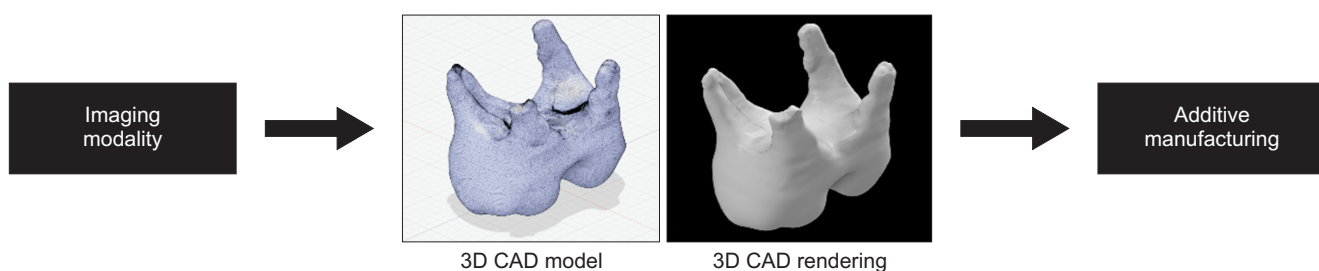


Fig. 1. The advent of additive manufacturing allows for the use of medical and research based imaging modalities to create three-dimensional (3D) computer aided design (CAD) models. These models can be rendered for visual enhancement and surgical simulation or the models can be converted to proper code for additive manufacturing into a graft, prototype or surgical model.

Laura Gaviria et al: Three-dimensional printing for craniomaxillofacial regeneration. *J Korean Assoc Oral Maxillofac Surg* 2017

parameters are needed to produce complex structures (e.g., interconnected porosity) with structural integrity, high quality and safety (e.g., sterility)^{12,14,22}. The following sections will describe the different types of materials used in CMF repair and approaches for 3D printing them.

1) Titanium

Titanium has a long history as a bone implant material because of its biocompatibility, strength to weight ratio and osteoconductive properties. In cranioplasty, titanium has been used in the form of sheets and meshes prefabricated using 3D printing techniques such as direct metal laser sintering¹⁵. Dental and CMF implants, plates and screws have been fabricated using titanium⁴ and although the use of this material has proven to be useful and clinically established, titanium implants cannot be replaced by ingrowing bone or function as a carrier for bioactive molecules^{14,18}.

2) Ceramics

Ceramics are commonly used in biomedical applications due to their high stiffness and bioactivity. Currently ceramic-based inks are available for direct 3D printing to fabricate patient specific bone grafts for dental and CMF repair applications^{4,24}. The most popular ceramics are calcium phosphates such as tri-calcium phosphates (TCP) and hydroxyapatite (HA) because of their excellent bioactivity, osteoconductivity, similarity to the mineral component of bone and bioresorptive properties^{12,16,17,19,22,34}. Previous studies have demonstrated their suitability for the build-up of 3D printed structures with resolutions of ~50 μm ¹⁶ as well as structures with controlled open pores that are capable of increasing osteoconduction *in vivo*¹². Also, evidence has shown the printability of a combination of TCP and bioactive glass which can be compositionally optimized for tailored biodegradation^{16,19}. Extensive research of 3D printing parameters such as powder packing, drop penetration, particle size, and calcium phosphate ratios has to be done for optimization of the 3D printed constructs^{17,24,34,35}.

3) Polymers

Blends of natural and synthetic polymeric biomaterial inks are adequate for printing scaffolds used in medical applications and can be customized for individual needs and applications in the CMF region²². In general, synthetic polymers are often poorly soluble in aqueous media, meaning that organic solvents must be used which raises concerns related to biocompatibility and large scale production of implants. Nonetheless, synthetic polymers are of great interest due to their

biocompatibility properties, ease of use, cost and degradation kinetics²⁴. The most used polymers in 3D printing for hard and soft tissues are polylactic acid (PLA), poly(caprolactone) (PCL), polyether ether ketone (PEEK)²⁴.

4) Composites

Although the initial 3D printing focused on pure materials, composite materials appear to be a most promising approach for the improvement and optimization of biomaterials at the engineering level²². The main goal of using composite inks is to enhance ink properties such as synthesis, printability, mechanics and bioactivity²⁴. Commercial 3D printers can be adapted for co-printing of polymer blends (polymer-based composites) and hydrogel-based composites²⁴. Other alternatives to improve mechanical and biological properties have been to add powdered ceramics as well as metals to pure polymers or polymer blends which can be printed using 3D printing nozzles^{17,22-24,34}.

III. Advantages and Limitations of 3D Printing Technology

3D printing offers outstanding possibilities in many aspects when compared to other methods, because it is more precise, faster, easily produced and cost-effective in a limited number of cases^{16,26}, eliminating highly specialized manual labor. 3D printing also offers advantages such as high versatility and capability to print complex designs^{20,27} using a large variety of biomaterials that can be printed individually or in combination^{18,21,22}.

Although industrial 3D printers, such as Stratasys Polyjet printers, have reached extremely high resolution (~16 μm) in the past few years^{4,16,21,23}, the use of 3D printing technology for implantable biomedical devices is still severely limited by available printable materials that cannot compete with traditional biomedical treatments. The main challenges are the use of processing methods required to work with materials that are not easily printed^{12,23,24,35} with the use of organic solvents and high processing temperatures which can harm and reduce the working life of 3D printers that are not specifically optimized for those very narrow uses^{23,35}. In summary, the main issues to be addressed in 3D printing of biomaterials are:

- The feasibility of low temperature 3D printing, especially for ceramic materials to make them more stable (control shrinkage) with the potential of incorporating biomolecules and polymers^{12,17,34}.

- The development of aqueous binder solutions used in scaffold fabrication to avoid the use of organic solvents that can compromise not only the biocompatibility of the scaffold but also the lifespan of the printer heads. This idea has been gaining attention because of its significant contribution to large-scale manufacturing^{12,17,34,35}.

- Achievement of high resolution and accurate porous interconnected structures with adequate mechanical and degradation properties. This approach can be optimized using composite biomaterial blends, as well as post processing treatments^{12,14,16,22-24}.

Overcoming the above mentioned technological limitations will finally lead to the incorporation of cells and growth factors/drugs to 3D printed scaffolds, since most of the currently used processing techniques cannot sustain the viability of cells and biomolecules after printing. This approach can tremendously impact the performance of the 3D printed constructs by balancing mechanical, biological, drug delivery and degradation properties^{12,20,22,30}. Future advancements in this field can be based on “multi-color” or “multi-component” printing, where each ink can be positioned on a precise location, offering the potential to simultaneously arrange multiple types of cells, deposit multiple extra cellular matrix materials, and exert point-to-point control over bioactive agents for biological tissue manufacturing²³. However, this approach only relies on the modification of current 3D printing machines and processing temperatures in order to maintain adequate conditions for cells and biomolecules²³. To date, some bio-printing technologies have been introduced and investigated in order to achieve this purpose²⁰, providing new insights into the future of complex tissue regeneration for bone, cartilage,

muscles, vessels, ligaments, tendons and nerves in the CMF complex²³. To such end, the following sections focus on specific treatments and approaches for regeneration of the periodontium and teeth within the oral cavity.

IV. Regeneration of the Periodontium

Periodontal disease afflicts approximately half of the population over thirty years of age in the United States. Genetic, environmental, dermatological and hematological factors all influence the high prevalence of this disease³⁶. Around 30% of the cases are characterized by moderate periodontitis with mild and severe cases about even. However, the number of patients with moderate to severe periodontitis increase to 64% after 65 years³⁷. This reduces the function of the periodontium (combined cementum, periodontal ligament [PDL] and alveolar bone) which is to secure the teeth to the mandible. In severe cases of periodontitis, the periodontium is destroyed and ultimately causing tooth loss^{38,39}. This destruction also leads to various complications and medical intervention, highlighting the need for a viable tissue regenerative approach.

1. The periodontium and strategies for regeneration

The complexity of tissue regeneration of the periodontium lie within the tissues of which it is comprised. The periodontal structure begins with the PDL which is unique in its shape and function. The web-like PDL connects the alveolar bone root to the cementum of the tooth providing tensile strength in a gap less than half of a millimeter³⁸ and support for mastication. Unlike other ligament attachments to bone throughout

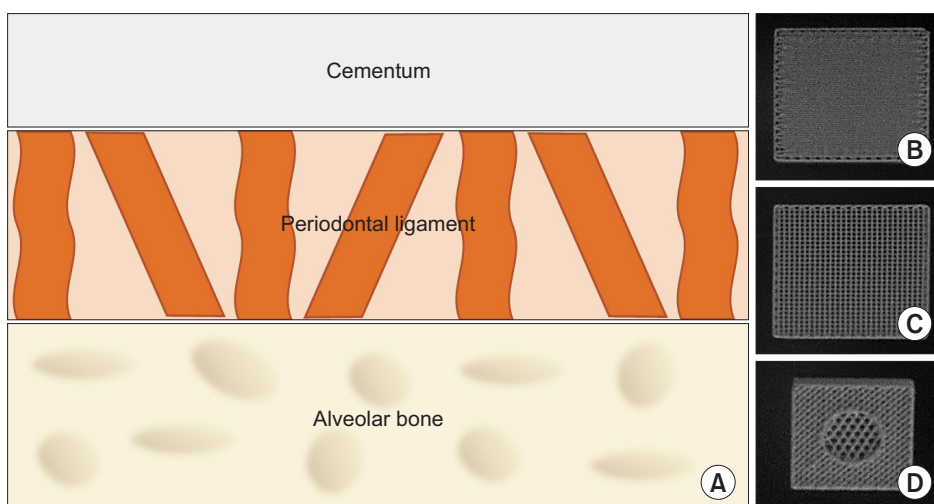


Fig. 2. The periodontium complex is comprised of cementum (A), the periodontal ligament and alveolar bone. These all have distinct porosity and strength. Additive manufacturing allows for different porosities and strengths (B, C) and the ability to create variations within the same grafts (D).

Laura Gaviria et al: Three-dimensional printing for craniomaxillofacial regeneration. J Korean Assoc Oral Maxillofac Surg 2017

the axial skeletal in which the ligament generally forms one insertion point on the bone, the PDL lines the entire surface with multiple small fibrous units being inserted at varying angles. Connective tissue and vascularization are intertwined with the PDL⁴⁰. Tissue regenerative approaches may seek to form a viable PDL structure, but most incorporate the entire periodontal complex or periodontium to combat the detachment of tissue caused by periodontitis⁴¹. The overall necessity for such approaches arises from the increasing concern of periodontitis described above and the lack of periodontium in current dental implants which will be discussed later in this review. Regenerative approaches for the periodontium have included growth factors, various cell types and materials that seek to provide adequate porosity and mechanics for the varying tissue types⁴². Moreover, additive manufacturing uses line spacing, line thickness and resolution to change mechanics and porosity.(Fig. 2) The advent of additive manufactur-

ing platforms such as the EnvisonTec 3D Bioplotter have broadened the capacity to 3D print a variety of biomaterials through ink development, and the continued reduction in cost of these platforms will make these technologies more readily available and should further expand the additive manufacturing tactics to periodontium regeneration. The following are current approaches using several materials, growth factors and cells.

2. Materials, manufacturing and cells in periodontium regeneration

Several studies performed for the regeneration of the periodontum are summarized in Table 1. A recent study created Mg-calcium-silicate cements with varying amounts of Mg which were seeded with PDL cells and evaluated for both odontogenesis and angiogenesis, which is a vital component

Table 1. An overview of various regenerative approaches discussed in this review and the diverse additive and other manufacturing techniques

Author	Journal	Periodontium regeneration		
		Synthesis technique	Tissue	Regenerative approach
Gerçek et al. ⁴⁵	<i>J Biomed Mater Res A</i> (2008)	Solvent/lyophilization	PDL	Used different PCL concentrations in tetrahydrofuran that formed microspheres after undergoing lyophilization and exhibited higher mechanical properties.
Oortgiesen et al. ⁴⁶	<i>Tissue Eng Part C Methods</i> (2012)	Gel substrate	PDL	Encapsulated PDL cells in collagen gels and evaluated under mechanical and chemical (enamel matrix derivative) stimulus.
Li et al. ⁴⁷	<i>Tissue Eng Part A</i> (2008)	Cell/substrate	Periodontium	Created dentin with transforming growth factor-β1 loaded Millipore filters <i>in vivo</i> . Then the filters were removed and PDL cells were seeded.
Park et al. ⁴⁸	<i>J Dent Res</i> (2014)	Directional freezing	Periodontal tissue	Placed dry ice at varying locations surrounding a paraffin tooth mold in a gelatin bath allowing for directional control of fibers.
Hasegawa et al. ⁵⁴	<i>Tissue Eng</i> (2005)	Temperature release	PDL	PDL cell sheets were created using thermosensitive PIPAAm to allow release of the sheets without using trypsin-EDTA.
Dan et al. ⁵⁵	<i>Biomaterials</i> (2014)	Melt electrospinning	Periodontal tissue	PCL with CaP coating scaffolds were implanted with either gingival, PDL or alveolar bone cell sheets.
Iwasaki et al. ⁵⁶	<i>Tissue Eng Part A</i> (2014)	Decellularization	Periodontal tissue	The decellularized amnion tissue was seeded with PDLSCs and assessed for cell viability with movement and surgery.
Lee et al. ⁶⁰	<i>Tissue Eng Part A</i> (2014)	3D printing	Periodontium	Three phase scaffolds (PCL with 10% HA) with different pores for the cementum, PDL and alveolar bone loaded with amelogenin, connective tissue growth factor and bone morphogenetic protein 2, respectively.
Pilipchuk et al. ⁶¹	<i>Adv Healthc Mater</i> (2016)	3D printing, patterning	Periodontium	Printed regions for bone (PCL with 5% HA) and patterned ligament (PCL) for cell alignment compared to salt leached scaffolds.
Ma et al. ⁶²	<i>Biofabrication</i> (2015)	Dropwise 3D printing	Periodontal tissue	Printed hydrogels with gradients of GelMA and PEG with encapsulated PDLSCs.
Rasperini et al. ⁶³	<i>J Dent Res</i> (2015)	3D printing, SLS	PDL/alveolar bone	Utilized computed tomography images to create a patient specific graft with SLS of PCL with 4% HA.

(PDL: periodontal ligament, PCL: poly(caprolactone), PIPAAm: poly(N-isopropylacrylamide), EDTA: ethylenediaminetetraacetic acid, CaP: calcium phosphate, PDLSCs: stem cells from the periodontal ligament, 3D: three-dimensional, HA: hydroxyapatite, GelMA: gelatin methacryloyl, PEG: polyethylene glycol, SLS: selective laser sintering)

Laura Gaviria et al: Three-dimensional printing for craniomaxillofacial regeneration. *J Korean Assoc Oral Maxillofac Surg* 2017

in the periodontal complex⁴³ co-existing with the PDL between the alveolar bone and cementum. It was discerned that higher Mg content provided higher angiogenic expression and may be an option for future studies⁴⁴. Another group lyophilized PCL in tetrahydrofuran to synthesize microspheres which could successfully maintain of PDL cells⁴⁵. PDL cells have also been incorporated into a collagen gel delivery system and stimulated via mechanical and chemical means. The unilateral loading alone increased alignment and cell number while the combination of mechanical stimulus with Emdogain (a protein-based stimulus for periodontal regeneration) did not produce improved results⁴⁶. In another study, PDL cells were cultured on dentin that was regenerated on readily available Millipore transfilters loaded with transforming growth factor- β 1 to ascertain the ability of dentin to regenerate PDL tissue⁴⁷. A unique approach to manufacturing periodontal scaffolds came from directional freezing followed by lyophilization. The approach used paraffin molds of the tooth and socket to form a gelatin periodontal complex which was frozen directionally by placing the ice at different regions surrounding the mold. This allowed for variation in the gelatin surface and the formation of a PDL template of fibers frozen in different directions which were lyophilized⁴⁸. Other manufacturing techniques used in PDL regeneration are electrospinning and melt electrospinning which provide random fibrous meshes as platforms for regeneration. These manufacturing techniques, materials and mechanical stimuli provide a solid base with which to model future studies. Moreover, these techniques combined with cells such as stem cells from the periodontal ligament (PDLSCs), alveolar bone stem cells (ABSCs), and dental pulp stem cells (DPSCs), and other primary cells⁴⁹⁻⁵¹, growth factors and enhanced spatiotemporal control using additive manufacturing could lead to a regenerative solution for the periodontal complex. Also, as discussed earlier, the importance of vascularization in the regenerative process is essential when postulating cell type and culture environment⁴³, and as such, PDLSCs have also been evaluated for angiogenic response. When seeded with endothelial cells (ECs), the PDLSCs increased the expression of vascular endothelial growth factor compared to ECs alone⁵². On collagen gels, PDLSCs can also differentiate towards an osteoblastic lineage to form alveolar bone⁵³ which connects with the PDL.

One tactic for employing the regenerative capacity of cells is to create cell sheets from cells of each tissue type. This can be achieved by releasing the cell sheet from poly(N-isopropylacrylamide) (PIPAAm) which changes hydrophilic-

ity at lower temperatures and allows the cells and extracellular matrix to detach without using trypsin⁵⁴. These sheets can be incorporated into porous scaffolds or electrospun meshes⁵⁵. One study evaluated cell sheets formed from PDL, alveolar bone and gingival cells on a PCL scaffold with melt electrospun bone and electrospun PDL sections. The alveolar and PDL cell sheets produced periodontal regeneration whereas the gingival cell sheet did not⁵⁵. PDLSC therapy has also been employed by seeding PDLSCs on decellularized amniotic membranes for transplantation⁵⁶. These cells can be combined with growth factors delivered in scaffolds to enhance regeneration. The main concern is being able to spatiotemporally control the delivery of cells and growth factors. Although the complexity of the periodontium and current techniques make this difficult⁴², additive manufacturing technology has the ability to improve the spatiotemporally control of tissue regeneration.

3. Additive manufacturing in periodontium regeneration

The continued advancement of additive manufacturing has allowed for printing of more materials and the printing of the same materials in conditions more relevant to tissue regeneration⁵⁷. This section gives an overview of the many advantages of additive manufacturing for periodontium regeneration. The first example is a PCL/HA scaffold of composite material manufactured in a layer-by-layer fashion by 3D printing using a 3D model created from laser scanning. Then, the scaffolds were infiltrated with growth factors such as stromal cell-derived factor-1 (SDF-1) and bone morphogenetic protein-7 (BMP-7)⁵⁸ in a collagen gel solution, showing significantly higher cell infiltration and angiogenesis⁵⁹. Other study printed PCL/HA composite scaffolds using the EnvisionTec 3D Bioplotter which is a pneumatic-based system that allows the user to vary parameters based on the solution viscosity. The scaffolds were triphasic in that the design changed mesh size for all three components of the periodontal complex with the alveolar bone and cementum having a smaller, stiffer mesh compared to the PDL. The scaffolds were loaded with poly(lactide-co-glycolide) (PLGA) microspheres with recombinant human amelogenin, connective tissue growth factor and bone morphogenetic protein 2 (BMP-2) in the cementum, PDL and alveolar bone sections respectively. *In vivo* evaluation with DPSCs found proper expression of bone and cementum tissues and alignment of collagen fibers in the PDL region⁶⁰. Other examples are the use of selective laser sintering to produce a PCL/HA scaffold with grooved pat-

tering for cell alignment⁶¹ compared to the previously discussed pneumatic approach, whereas another additive manufacturing technique uses a dropwise gel printing method with cell encapsulation. Using dropwise gel printing technique, one study printed PDLSCs dropwise in gelatin methacryloyl/polyethylene glycol (PEG) hydrogels with a gradient of the two hydrogel components (ratios for 0:5 to 5:0 across a well plate) and reported that the lower ratios of PEG increased cell viability, area and proliferation⁶².

Some additive manufacturing approaches have moved beyond the periodontium to include a layer of native dentin to determine if the regenerated periodontium could form a functional junction with the dentin of the tooth. The use of a native tissue also has the capacity to reduce immunogenic response once implanted. Studies have employed this dentin technique leading up to and following the first human trial of a 3D printed periodontium which was completed in 2015 with a PCL-based scaffold with specific bone and PDL sections and a burst release of platelet-derived growth factor (PDGF). The implant was removed at 12 months showing that the long PCL degradation timeline was not advantageous and that other materials, degradation profiles and inductive factors should be explored⁶³. Moreover, additive manufacturing approaches for the regeneration of the periodontium and teeth combined can be very beneficial in this field. Next section of this review will discuss those approaches.

V. Regeneration of Teeth

The tooth is a complex organ formed by a variety soft (dental pulp) and hard tissues (dentin, enamel, and cementum)⁶⁴ which, in conjunction, maintain the physiological and biological environment³⁹. Enamel cannot regenerate itself in an adult tooth⁶⁵. If eroded, enamel can leave the tooth exposed and may lead to the need for tooth replacement with an implant or graft. These options for adult patients stress the importance of dental care and the ability to intervene with periodontium focused tissue regeneration as discussed above prior to the need for implants.

1. Dental implants, imaging and additive manufacturing

Dental implants are usually designed to allow for osteointegration and increased aesthetics. The process of implantation of synthetic implants takes months, with time taken for metal post integration after bone grafting and prior to final insertion of the abutment and synthetic crown. However, this

standard implant does not provide or seek to replace the PDL. The abutment screw is designed to create a preload that often determines the success of the implant through load sharing and osteointegration. If the post does not integrate into the alveolar bone, the implant can loosen. Nonetheless, preloading of the screw to certain values and subsequent loading after initial positioning has been shown to increase success⁶⁶.

New imaging techniques such as cone-beam CT and 3D modeling can be utilized not only for manufacturing scaffolds, but also for prototyping and preparing the implant site. The surgeon can use a stereolithographic additive (SLA) manufacturing technique to make a tooth prototype and shape the alveolar bone to fit the tooth that is to be transplanted. This reduces the transplantation time and helps maintain the vasculature and cells in the tooth⁶⁷. The prototypes can be directly printed through SLA manufacturing or resin cast into a 3D printed wax negative⁶⁸. These techniques provide an outlook on what additive manufacturing can offer in tooth regeneration. These same strategies can be used to take images of the teeth of a specific patient and 3D print a tooth to those exact specifications. However, this would not restore function because the wax and resin prints currently used cannot form a functional replacement tooth. However, with the advancement of strategies to print a tooth with biomaterials, growth factors and cells could harness this approach and create a functional solution. Additive manufacturing approaches towards tooth regeneration are explored below.

2. Additive manufacturing in tooth regeneration

Currently, tooth 3D printing is performed by different 3D printing technologies and extrusion methods^{69,70}. Different biomaterials such as collagen sponge^{71,72}, agarose⁷³, alginate⁷⁴, hyaluronan-chondroitin copolymers⁷⁵, poly-glycolic acid (PGA), PLA^{76,77}, and fibrin⁷⁸ have been paired with dental stem cells to regenerate different components of teeth including dental pulp⁷⁹⁻⁸¹, dentin^{82,83}, crown^{71,84} and roots⁸⁵⁻⁸⁷. From the wide variety of materials, ceramics such as HA and TCP are obvious candidates for regeneration of the tooth, alveolar bone complex due to their known osteoconductive properties⁸⁸.

3D printing techniques can manufacture scaffolds in the exact shape and size of the missing tooth using imaging the contralateral existing tooth^{18,89,90}. Other materials such as 3D printed alumina coated with HA⁷⁰, silica- β -TCP, zinc oxide- β -TCP⁹¹ and printable composites such as PLGA- β -TCP⁶⁹, as well as PCL- β -TCP⁹² and PCL-HA⁵⁹ have also been evalu-

ated for 3D printed regeneration of the tooth/alveolar bone complex. In 2010, Kim et al.⁵⁹ were the first team to demonstrate tissue ingrowth (including PDL) in an anatomically correct 3D bioprinted tooth scaffolds *in vivo*.

The ultimate goal in the development of 3D printed tooth scaffolds would be the incorporation of stem cells since this is an area that attracts great interest from the regenerative medicine viewpoint⁹³⁻⁹⁵. The ability of stem cells to differentiate into different cell types makes them a viable candidate for therapies that could result in the regeneration of the different tissues that form the tooth complex^{96,97}. Different types of stem cells have been investigated for their potential in tooth regeneration. DPSCs, as mentioned in periodontium regeneration, have also been identified to be capable of forming a structure similar to dentin lined by odontoblast-like cells surrounding a tissue comparable to dental pulp⁹⁸. Stem cells from human exfoliated deciduous teeth (SHEDs) have the ability to differentiate into odontoblasts, osteoblasts and adipocytes⁹⁹ and are easily accessible¹⁰⁰. When compared to DPSCs, SHEDs were shown to have a higher proliferation rate and differentiation capacity *in vitro* as well as a potentially higher mineralization capacity¹⁰¹. In addition, stem cells from apical papilla (SCAPS) have also been isolated from extracted wisdom teeth displaying greater potential for proliferation, stemness, and dentin regeneration than DPSCs¹⁰².

Since embryonic stem cell research has raised ethical concerns¹⁰³, most efforts have been focused on differentiating stem cells obtained from adult tissues or inducing embryonic-like pluripotency on other cells. The combinational use of these cell types together with the above additive manufacturing techniques comprised of osteoconductive biomaterials, medical imaging and 3D modelling has the potential to produce a patient specific tissue regenerative approach for teeth.

VI. Conclusion

CMF defects caused by disease, surgery or trauma are complex in nature and involve repair of many different tissue types with unique properties and intricate geometries. Therefore, CMF surgery not only represents a challenge for CMF surgeons, but it also poses a multifaceted design problem to fabricate a complex, 3D biomedical tissue regenerative alternative to current treatments²³. In recent years various tissue engineering approaches for CMF repair have been explored. Traditionally, the biomedical field has relied on manually fabricated scaffolds for hard and soft tissue. However, recent developments have adapted 3D printing into an increasingly

common technique to fabricate scaffolds and devices for CMF applications due to its potential to provide patient-specific designs, high structural complexity, and relatively rapid, fully-automated fabrication at a low-cost^{24,27}. Moreover, the long-term goal of 3D printing in tissue engineering will be to develop printable biomaterial inks capable of creating safe and reproducible scaffolds with tunable mechanical, biological and degradation properties^{22,23,34}. To achieve that, 3D printers need to continue to be re-designed with the specific capabilities needed for multi-component printing of biomaterials, viable cells and biomolecules in order to mimic the physiological environment and enhance tissue repair^{4,22-24}.

As shown in this review, there have been strides in CMF repair, especially in periodontium and tooth regeneration in large part due to the advancement in additive manufacturing techniques. The periodontium scaffold combined with native dentin slices⁶³ gives an outlook of the potential combination of multiple tissue additive manufacturing strategies to regenerate complex defects with many tissue types. Although, 3D printing holds great overall promise due to its diverse applicability in routine and complex cases of dental and CMF surgery and planning^{20,27}, there are still many technical challenges to overcome before it can be recognized as a common biofabrication technique in medicine^{24,26,29,104}.

Conflict of Interest

No potential conflict of interest relevant to this article was reported.

ORCID

Laura Gaviria, <http://orcid.org/0000-0003-3495-8762>

Joseph J. Pearson, <http://orcid.org/0000-0003-0054-5732>

Sergio A. Montelongo, <http://orcid.org/0000-0001-8923-0096>

Teja Guda, <http://orcid.org/0000-0002-3218-2916>

Joo L. Ong, <http://orcid.org/0000-0003-3330-2390>

References

1. Gadre KS, Halli R, Joshi S, Ramanojam S, Gadre PK, Kunchur R, et al. Incidence and pattern of cranio-maxillofacial injuries: a 22 year retrospective analysis of cases operated at major trauma hospitals/centres in Pune, India. *J Maxillofac Oral Surg* 2013;12:372-8.
2. Henderson R. Maxillofacial injuries [Internet]. Leeds, UK: Patient, 2014 [cited 2017 Jun 5]. Available from: patient.info/doctor/maxillofacial-injuries.

3. Gassner R, Tuli T, Hächl O, Rudisch A, Ulmer H. Cranio-maxillofacial trauma: a 10 year review of 9,543 cases with 21,067 injuries. *J Craniomaxillofac Surg* 2003;31:51-61.
4. Hikita A, Chung UI, Hoshi K, Takato T. Bone regenerative medicine in oral and maxillofacial region using a three-dimensional printer. *Tissue Eng Part A* 2017;23:515-21.
5. Lew TA, Walker JA, Wenke JC, Blackburne LH, Hale RG. Characterization of craniomaxillofacial battle injuries sustained by United States service members in the current conflicts of Iraq and Afghanistan. *J Oral Maxillofac Surg* 2010;68:3-7.
6. Hale RG, Lew T, Wenke JC. Craniomaxillofacial battle injuries: injury patterns, conventional treatment limitations and direction of future research. *Singapore Dent J* 2010;31:1-8.
7. Chan RK, Siller-Jackson A, Verrett AJ, Wu J, Hale RG. Ten years of war: a characterization of craniomaxillofacial injuries incurred during operations Enduring Freedom and Iraqi Freedom. *J Trauma Acute Care Surg* 2012;73(6 Suppl 5):S453-8.
8. Brown Baer PR, Wenke JC, Thomas SJ, Hale CR. Investigation of severe craniomaxillofacial battle injuries sustained by U.S. service members: a case series. *Craniomaxillofac Trauma Reconstr* 2012;5:243-52.
9. Carano RA, Filvaroff EH. Angiogenesis and bone repair. *Drug Discov Today* 2003;8:980-9.
10. Alsberg E, Hill EE, Mooney DJ. Craniofacial tissue engineering. *Crit Rev Oral Biol Med* 2001;12:64-75.
11. Kraft A, Abermann E, Stigler R, Zsifkovits C, Pedross F, Kloss F, et al. Craniomaxillofacial trauma: synopsis of 14,654 cases with 35,129 injuries in 15 years. *Craniomaxillofac Trauma Reconstr* 2012;5:41-50.
12. Bose S, Vahabzadeh S, Bandyopadhyay A. Bone tissue engineering using 3D printing. *Materials Today* 2013;16:496-504.
13. Mehta M, Schmidt-Bleek K, Duda GN, Mooney DJ. Biomaterial delivery of morphogens to mimic the natural healing cascade in bone. *Adv Drug Deliv Rev* 2012;64:1257-76.
14. Klammert U, Gbureck U, Vorndran E, Rödiger J, Meyer-Marcotty P, Kübler AC. 3D powder printed calcium phosphate implants for reconstruction of cranial and maxillofacial defects. *J Craniomaxillofac Surg* 2010;38:565-70.
15. Parthasarathy J. 3D modeling, custom implants and its future perspectives in craniofacial surgery. *Ann Maxillofac Surg* 2014;4:9-18.
16. Bergmann C, Lindner M, Zhang W, Koczur K, Kirsten A, Telle R, et al. 3D printing of bone substitute implants using calcium phosphate and bioactive glasses. *J Eur Ceram Soc* 2010;30:2563-7.
17. Inzana JA, Olvera D, Fuller SM, Kelly JP, Graeve OA, Schwarz EM, et al. 3D printing of composite calcium phosphate and collagen scaffolds for bone regeneration. *Biomaterials* 2014;35:4026-34.
18. Cohen A, Laviv A, Berman P, Nashef R, Abu-Tair J. Mandibular reconstruction using stereolithographic 3-dimensional printing modeling technology. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2009;108:661-6.
19. Gross BC, Erkal JL, Lockwood SY, Chen C, Spence DM. Evaluation of 3D printing and its potential impact on biotechnology and the chemical sciences. *Anal Chem* 2014;86:3240-53.
20. Farré-Guasch E, Wolff J, Helder MN, Schulten EA, Forouzanfar T, Klein-Nulend J. Application of additive manufacturing in oral and maxillofacial surgery. *J Oral Maxillofac Surg* 2015;73:2408-18.
21. Xiao K, Zardawi F, van Noort R, Yates JM. Developing a 3D colour image reproduction system for additive manufacturing of facial prostheses. *Int J Adv Manuf Technol* 2014;70:2043-9.
22. Butscher A, Bohner M, Hofmann S, Gauckler L, Müller R. Structural and material approaches to bone tissue engineering in powder-based three-dimensional printing. *Acta Biomater* 2011;7:907-20.
23. Chia HN, Wu BM. Recent advances in 3D printing of biomaterials. *J Biol Eng* 2015;9:4.
24. Guvendiren M, Molde J, Soares RM, Kohn J. Designing biomaterials for 3D printing. *ACS Biomater Sci Eng* 2016;2:1679-93.
25. Liu YF, Xu LW, Zhu HY, Liu SS. Technical procedures for template-guided surgery for mandibular reconstruction based on digital design and manufacturing. *Biomed Eng Online* 2014;13:63.
26. Schwam ZG, Chang MT, Barnes MA, Paskhover B. Applications of 3-dimensional printing in facial plastic surgery. *J Oral Maxillofac Surg* 2016;74:427-8.
27. Dawood A, Marti Marti B, Sauret-Jackson V, Darwood A. 3D printing in dentistry. *Br Dent J* 2015;219:521-9.
28. Malik HH, Darwood AR, Shaunak S, Kulatilake P, El-Hilly AA, Mulki O, et al. Three-dimensional printing in surgery: a review of current surgical applications. *J Surg Res* 2015;199:512-22.
29. Rengier F, Mehndiratta A, von Tengg-Kobligh H, Zechmann CM, Unterhinninghofen R, Kauczor HU, et al. 3D printing based on imaging data: review of medical applications. *Int J Comput Assist Radiol Surg* 2010;5:335-41.
30. Cillo JE Jr, Basi D, Peacock Z, Aghaloo T, Bouloux G, Dodson T, et al. Proceedings of the American Association of Oral and Maxillofacial Surgeons 2015 Research Summit. *J Oral Maxillofac Surg* 2016;74:429-37.
31. Velasco I, Vahdani S, Ramos H, Guzman J. Clinical application of desktop three-dimensional printing technology in ablative and reconstructive maxillofacial surgery. *Oral Surg Oral Med Oral Pathol Oral Radiol* 2017;123:e23-4.
32. Thomas DJ, Azmi MABM, Tehrani Z. 3D additive manufacture of oral and maxillofacial surgical models for preoperative planning. *Int J Adv Manuf Technol* 2014;71:1643-51.
33. Leukers B, Gülkan H, Irsen SH, Milz S, Tille C, Schieker M, et al. Hydroxyapatite scaffolds for bone tissue engineering made by 3D printing. *J Mater Sci Mater Med* 2005;16:1121-4.
34. Khalyfa A, Vogt S, Weisser J, Grimm G, Rechtenbach A, Meyer W, et al. Development of a new calcium phosphate powder-binder system for the 3D printing of patient specific implants. *J Mater Sci Mater Med* 2007;18:909-16.
35. Zhou Z, Buchanan F, Mitchell C, Dunne N. Printability of calcium phosphate: calcium sulfate powders for the application of tissue engineered bone scaffolds using the 3D printing technique. *Mater Sci Eng C Mater Biol Appl* 2014;38:1-10.
36. Pihlstrom BL, Michalowicz BS, Johnson NW. Periodontal diseases. *Lancet* 2005;366:1809-20.
37. Eke PI, Dye BA, Wei L, Thornton-Evans GO, Genco RJ. Prevalence of periodontitis in adults in the United States: 2009 and 2010. *J Dent Res* 2012;91:914-20.
38. Wang J, Zhang R, Shen Y, Xu C, Qi S, Lu L, et al. Recent advances in cell sheet technology for periodontal regeneration. *Curr Stem Cell Res Ther* 2014;9:162-73.
39. Tatullo M, Marrelli M, Shakesheff KM, White LJ. Dental pulp stem cells: function, isolation and applications in regenerative medicine. *J Tissue Eng Regen Med* 2015;9:1205-16.
40. Nanci A, Bosshardt DD. Structure of periodontal tissues in health and disease. *Periodontol* 2000 2006;40:11-28.
41. Kim JH, Park CH, Perez RA, Lee HY, Jang JH, Lee HH, et al. Advanced biomatrix designs for regenerative therapy of periodontal tissues. *J Dent Res* 2014;93:1203-11.
42. Ivanovski S, Vaquette C, Gronthos S, Huttmacher DW, Bartold PM. Multiphasic scaffolds for periodontal tissue engineering. *J Dent Res* 2014;93:1212-21.
43. Jakab K, Norotte C, Marga F, Murphy K, Vunjak-Novakovic G, Forgacs G. Tissue engineering by self-assembly and bio-printing of living cells. *Biofabrication* 2010;2:022001.
44. Chen YW, Hsu TT, Wang K, Shie MY. Preparation of the fast setting and degrading Ca-Si-Mg cement with both odontogenesis and angiogenesis differentiation of human periodontal ligament cells. *Mater Sci Eng C Mater Biol Appl* 2016;60:374-83.
45. Gerçek I, Tigli RS, Gümüşderelioglu M. A novel scaffold based on formation and agglomeration of PCL microbeads by freeze-drying. *J Biomed Mater Res A* 2008;86:1012-22.

46. Oortgiesen DA, Yu N, Bronckers AL, Yang F, Walboomers XF, Jansen JA. A three-dimensional cell culture model to study the mechano-biological behavior in periodontal ligament regeneration. *Tissue Eng Part C Methods* 2012;18:81-9.
47. Li Y, Jin F, Du Y, Ma Z, Li F, Wu G, et al. Cementum and periodontal ligament-like tissue formation induced using bioengineered dentin. *Tissue Eng Part A* 2008;14:1731-42.
48. Park CH, Kim KH, Rios HF, Lee YM, Giannobile WV, Seol YJ. Spatiotemporally controlled microchannels of periodontal mimic scaffolds. *J Dent Res* 2014;93:1304-12.
49. Eleuterio E, Trubiani O, Sulpizio M, Di Giuseppe F, Pierdomenico L, Marchisio M, et al. Proteome of human stem cells from periodontal ligament and dental pulp. *PLoS One* 2013;8:e71101.
50. Horst OV, Chavez MG, Jheon AH, Desai T, Klein OD. Stem cell and biomaterials research in dental tissue engineering and regeneration. *Dent Clin North Am* 2012;56:495-520.
51. Liu B, Song YW, Jin L, Wang ZJ, Pu DY, Lin SQ, et al. Silk structure and degradation. *Colloids Surf B Biointerfaces* 2015;131:122-8.
52. Yeasmin S, Ceccarelli J, Vigen M, Carrion B, Putnam AJ, Tarle SA, et al. Stem cells derived from tooth periodontal ligament enhance functional angiogenesis by endothelial cells. *Tissue Eng Part A* 2014;20:1188-96.
53. Alves LB, Marigueta VC, Grisi MF, Souza SL, Novaes Junior AB, Taba Junior M, et al. Expression of osteoblastic phenotype in periodontal ligament fibroblasts cultured in three-dimensional collagen gel. *J Appl Oral Sci* 2015;23:206-14.
54. Hasegawa M, Yamato M, Kikuchi A, Okano T, Ishikawa I. Human periodontal ligament cell sheets can regenerate periodontal ligament tissue in an athymic rat model. *Tissue Eng* 2005;11:469-78.
55. Dan H, Vaquette C, Fisher AG, Hamlet SM, Xiao Y, Huttmacher DW, et al. The influence of cellular source on periodontal regeneration using calcium phosphate coated polycaprolactone scaffold supported cell sheets. *Biomaterials* 2014;35:113-22.
56. Iwasaki K, Komaki M, Yokoyama N, Tanaka Y, Taki A, Honda I, et al. Periodontal regeneration using periodontal ligament stem cell-transferred amnion. *Tissue Eng Part A* 2014;20:693-704.
57. Li X, Cui R, Sun L, Aifantis KE, Fan Y, Feng Q, et al. 3D-printed biopolymers for tissue engineering application. *Int J Polym Sci* 2014;2014:1-13.
58. Li J, He L, Zhou C, Zhou Y, Bai Y, Lee FY, et al. 3D printing for regenerative medicine: from bench to bedside. *MRS Bull* 2015;40:145-54.
59. Kim K, Lee CH, Kim BK, Mao JJ. Anatomically shaped tooth and periodontal regeneration by cell homing. *J Dent Res* 2010;89:842-7.
60. Lee CH, Hajibandeh J, Suzuki T, Fan A, Shang P, Mao JJ. Three-dimensional printed multiphase scaffolds for regeneration of periodontium complex. *Tissue Eng Part A* 2014;20:1342-51.
61. Pilipchuk SP, Monje A, Jiao Y, Hao J, Kruger L, Flanagan CL, et al. Integration of 3D printed and micropatterned polycaprolactone scaffolds for guidance of oriented collagenous tissue formation in vivo. *Adv Healthc Mater* 2016;5:676-87.
62. Ma Y, Ji Y, Huang G, Ling K, Zhang X, Xu F. Bioprinting 3D cell-laden hydrogel microarray for screening human periodontal ligament stem cell response to extracellular matrix. *Biofabrication* 2015;7:044105.
63. Rasperini G, Pilipchuk SP, Flanagan CL, Park CH, Pagni G, Hollister SJ, et al. 3D-printed bioresorbable scaffold for periodontal repair. *J Dent Res* 2015;94(Suppl):153S-7S.
64. Yildirim S, Fu SY, Kim K, Zhou H, Lee CH, Li A, et al. Tooth regeneration: a revolution in stomatology and evolution in regenerative medicine. *Int J Oral Sci* 2011;3:107-16.
65. Kwak SY, Litman A, Margolis HC, Yamakoshi Y, Simmer JP. Biomimetic enamel regeneration mediated by leucine-rich amelogenin peptide. *J Dent Res* 2017;96:524-30.
66. Siamos G, Winkler S, Boberick KG. Relationship between implant preload and screw loosening on implant-supported prostheses. *J Oral Implantol* 2002;28:67-73.
67. Cross D, El-Angbawi A, McLaughlin P, Keightley A, Brocklebank L, Whitters J, et al. Developments in autotransplantation of teeth. *Surgeon* 2013;11:49-55.
68. Kato A, Ohno N. Construction of three-dimensional tooth model by micro-computed tomography and application for data sharing. *Clin Oral Investig* 2009;13:43-6.
69. Li J, Zhang L, Lv S, Li S, Wang N, Zhang Z. Fabrication of individual scaffolds based on a patient-specific alveolar bone defect model. *J Biotechnol* 2011;151:87-93.
70. Bose S, Darsell J, Hosick HL, Yang L, Sarkar DK, Bandyopadhyay A. Processing and characterization of porous alumina scaffolds. *J Mater Sci Mater Med* 2002;13:23-8.
71. Sumita Y, Honda MJ, Ohara T, Tsuchiya S, Sagara H, Kagami H, et al. Performance of collagen sponge as a 3-D scaffold for tooth-tissue engineering. *Biomaterials* 2006;27:3238-48.
72. Honda MJ, Tsuchiya S, Sumita Y, Sagara H, Ueda M. The sequential seeding of epithelial and mesenchymal cells for tissue-engineered tooth regeneration. *Biomaterials* 2007;28:680-9.
73. Sloan AJ, Rutherford RB, Smith AJ. Stimulation of the rat dentine-pulp complex by bone morphogenetic protein-7 in vitro. *Arch Oral Biol* 2000;45:173-7.
74. Dobie K, Smith G, Sloan AJ, Smith AJ. Effects of alginate hydrogels and TGF-beta 1 on human dental pulp repair in vitro. *Connect Tissue Res* 2002;43:387-90.
75. Kuo TF, Huang AT, Chang HH, Lin FH, Chen ST, Chen RS, et al. Regeneration of dentin-pulp complex with cementum and periodontal ligament formation using dental bud cells in gelatin-chondroitin-hyaluronan tri-copolymer scaffold in swine. *J Biomed Mater Res A* 2008;86:1062-8.
76. Young CS, Terada S, Vacanti JP, Honda M, Bartlett JD, Yelick PC. Tissue engineering of complex tooth structures on biodegradable polymer scaffolds. *J Dent Res* 2002;81:695-700.
77. Duailibi MT, Duailibi SE, Young CS, Bartlett JD, Vacanti JP, Yelick PC. Bioengineered teeth from cultured rat tooth bud cells. *J Dent Res* 2004;83:523-8.
78. Anitua E, Alkhraisat MH, Orive G. Perspectives and challenges in regenerative medicine using plasma rich in growth factors. *J Control Release* 2012;157:29-38.
79. Iohara K, Nakashima M, Ito M, Ishikawa M, Nakasima A, Akamine A. Dentin regeneration by dental pulp stem cell therapy with recombinant human bone morphogenetic protein 2. *J Dent Res* 2004;83:590-5.
80. Huang GT, Yamaza T, Shea LD, Djouad F, Kuhn NZ, Tuan RS, et al. Stem/progenitor cell-mediated de novo regeneration of dental pulp with newly deposited continuous layer of dentin in an in vivo model. *Tissue Eng Part A* 2010;16:605-15.
81. Janjić K, Cvikić B, Moritz A, Agis H. Dental pulp regeneration. *Int J Stomat Occ Med* 2016;8(Suppl 1):1-9.
82. Guo W, He Y, Zhang X, Lu W, Wang C, Yu H, et al. The use of dentin matrix scaffold and dental follicle cells for dentin regeneration. *Biomaterials* 2009;30:6708-23.
83. Li R, Guo W, Yang B, Guo L, Sheng L, Chen G, et al. Human treated dentin matrix as a natural scaffold for complete human dentin tissue regeneration. *Biomaterials* 2011;32:4525-38.
84. Hu B, Nadiri A, Kuchler-Bopp S, Perrin-Schmitt F, Peters H, Lesot H. Tissue engineering of tooth crown, root, and periodontium. *Tissue Eng* 2006;12:2069-75.
85. Yang B, Chen G, Li J, Zou Q, Xie D, Chen Y, et al. Tooth root regeneration using dental follicle cell sheets in combination with a dentin matrix-based scaffold. *Biomaterials* 2012;33:2449-61.
86. Wei F, Song T, Ding G, Xu J, Liu Y, Liu D, et al. Functional tooth restoration by allogeneic mesenchymal stem cell-based bio-root regeneration in swine. *Stem Cells Dev* 2013;22:1752-62.
87. Rosa V, Zhang Z, Grande RH, Nör JE. Dental pulp tissue engineering in full-length human root canals. *J Dent Res* 2013;92:970-5.

88. Guda T, Appleford M, Oh S, Ong JL. A cellular perspective to bio-ceramic scaffolds for bone tissue engineering: the state of the art. *Curr Top Med Chem* 2008;8:290-9.
89. McMenamin PG, Quayle MR, McHenry CR, Adams JW. The production of anatomical teaching resources using three-dimensional (3D) printing technology. *Anat Sci Educ* 2014;7:479-86.
90. Obregon F, Vaquette C, Ivanovski S, Hutmacher DW, Bertassoni LE. Three-dimensional bioprinting for regenerative dentistry and craniofacial tissue engineering. *J Dent Res* 2015;94(9 Suppl):143S-52S.
91. Fielding GA, Bandyopadhyay A, Bose S. Effects of silica and zinc oxide doping on mechanical and biological properties of 3D printed tricalcium phosphate tissue engineering scaffolds. *Dent Mater* 2012;28:113-22.
92. Vaquette C, Fan W, Xiao Y, Hamlet S, Hutmacher DW, Ivanovski S. A biphasic scaffold design combined with cell sheet technology for simultaneous regeneration of alveolar bone/periodontal ligament complex. *Biomaterials* 2012;33:5560-73.
93. Gimble JM, Katz AJ, Bunnell BA. Adipose-derived stem cells for regenerative medicine. *Circ Res* 2007;100:1249-60.
94. Caplan AI. Adult mesenchymal stem cells for tissue engineering versus regenerative medicine. *J Cell Physiol* 2007;213:341-7.
95. Yu J, Vodyanik MA, Smuga-Otto K, Antosiewicz-Bourget J, Frane JL, Tian S, et al. Induced pluripotent stem cell lines derived from human somatic cells. *Science* 2007;318:1917-20.
96. Yan M, Yu Y, Zhang G, Tang C, Yu J. A journey from dental pulp stem cells to a bio-tooth. *Stem Cell Rev* 2011;7:161-71.
97. Otsu K, Kumakami-Sakano M, Fujiwara N, Kikuchi K, Keller L, Lesot H, et al. Stem cell sources for tooth regeneration: current status and future prospects. *Front Physiol* 2014;5:36.
98. Gronthos S, Mankani M, Brahimi J, Robey PG, Shi S. Postnatal human dental pulp stem cells (DPSCs) in vitro and in vivo. *Proc Natl Acad Sci U S A* 2000;97:13625-30.
99. Miura M, Gronthos S, Zhao M, Lu B, Fisher LW, Robey PG, et al. SHED: stem cells from human exfoliated deciduous teeth. *Proc Natl Acad Sci U S A* 2003;100:5807-12.
100. Ma L, Makino Y, Yamaza H, Akiyama K, Hoshino Y, Song G, et al. Cryopreserved dental pulp tissues of exfoliated deciduous teeth is a feasible stem cell resource for regenerative medicine. *PLoS One* 2012;7:e51777.
101. Wang X, Sha XJ, Li GH, Yang FS, Ji K, Wen LY, et al. Comparative characterization of stem cells from human exfoliated deciduous teeth and dental pulp stem cells. *Arch Oral Biol* 2012;57:1231-40.
102. Sonoyama W, Liu Y, Fang D, Yamaza T, Seo BM, Zhang C, et al. Mesenchymal stem cell-mediated functional tooth regeneration in swine. *PLoS One* 2006;1:e79.
103. de Wert G, Mummery C. Human embryonic stem cells: research, ethics and policy. *Hum Reprod* 2003;18:672-82.
104. Michalski MH, Ross JS. The shape of things to come: 3D printing in medicine. *JAMA* 2014;312:2213-4.