# The first record of nine bacterial species belonging to the phylum *Proteobacteria* in Korea

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As part of a larger study with the aim to discover indigenous prokaryotic species in Korea, nine bacterial strains were isolated and assigned to the phylum *Proteobacteria* in 2016. High 16S rRNA gene sequence similarity (>98.5%) and formation of a robust phylogenetic clades with known species indicated that each strain belongs to an independent and predefined bacterial species. This is the first report of these nine species in Korea: two strains of the *Methylobacterium*, two strains of the *Microvirga*, one strain of the *Pantoea*, and four strains of the *Psychrobacter*, all within the *Proteobacteria*. Gram reaction, colony and cell morphology, basic biochemical characteristics, and isolation sources are also described in the species description section.

Keywords: 16S rRNA, bacterial diversity, Proteobacteria, unreported species

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# **INTRODUCTION**

In 2016, we collected diverse soil samples and isolated unrecorded bacterial species in Korea. The identified bacterial species belonged to the phylum *Proteobacteria*. This report focuses on the isolation and description of unrecorded radiation-resistant species belonging to the phylum *Proteobacteria*.

Proteobacteria were established by Carl Woese (1987) and are one of the major phyla of Gram-negative bacteria. To date, the phylum Proteobacteria is known to be comprised of class Alphaproteobacteria, Betaproteobacteria, Gammaproteobacteria, Deltaproteobacteria, Epsilonproteobacteria, Zetaproteobacteria and Oligoflexia (List of Prokaryotic names with Standing in Nomenclature (LPSN); http://www.bacterio.net/-classifphyla.html #proteobacteria).

The *Alphaproteobacteria* with in the phylum *Proteobacteria* are a diverse class of organisms with many important biological roles including various metabolic strategies such as photosynthesis and nitrogen fixation. Moreover, they frequently adopt an intracellular life history as plant mutualists (Williams *et al.*, 2007).

*Gammaproteobacteria* include the several ecologically, and scientifically important groups of bacteria like

*Escherichia coli*, *Vibrio cholera* and *Salmonella* sp.. These bacteria play an important role as human pathogens. Although *Gammaproteobacteria* has only the taxonomic rank of class within the phylum *Proteobacteria*, it is richer in genera (~250) than all bacterial phyla except *Firmicutes* (Williams *et al.*, 2010).

This report focuses on the description of bacterial species belonging to the *Proteobacteria* that are new records for Korea. This study provides the first reports in Korea of nine bacterial species belonging to three families of three orders in the *Proteobacteria*.

# MATERIALS AND METHODS

Various soil samples that isolated from Jeju were suspended on distilled water and serially diluted. R2A, TSA and MA agar media were inoculated with aliquots and the plates were incubated at 25°C for 3 days (Table 1). The designated strain IDs, sources, culture media, and incubation conditions are summarized in Table 1. All strains were purified as single colonies and stored as a 20 % glycerol suspension at -80°C as well as lyophilized ampoules.

Colony morphology and cell size of the strains were

Table 1. 16S rRNA gene sequence similarity, isolation source, medium, and incubation conditions of unrecorded strains belonging to the genus *Methylobacterium* 

Strain ID	Most closely related species	Similarity (%)	Isolation source	Medium	Incubation conditions
15J10-8T5	Methylobacterium hispanicum	99.9	Soil of Jeju	R2A	25°C, 3d
KSM3-9	Methylobacterium soli	98.8	Soil of Jeju	R2A	25°C, 3d



**Fig. 1.** Transmission electron micrographs of the strains isolated in this study. Strains: 1, 15J10-8T5; 2, KSM3-9; 3, 15J8-3; 4, 15J7-3T5; 5, 15J17K; 6, 15ML1C; 7, 216MGT3; 8, 216MGM6; 9, 16MFM4.

observed on R2A agar after cells were grown for 3 days at 25°C by using transmission electron microscopy (LIBRA 120, Carl Zeiss). Transmission electron micrographs of the strains are shown in Fig. 1. Gram reaction was performed according to the classic Gram procedure described by Doetsch (1981). Biochemical characteristics were tested using Biolog Microstation with the GEN III microplate system. A single colony was selected and emulsified into 'inoculating fluid A' (Biolog) for subsequent inoculation on to the MicroPlate test plate (Biolog). More fastidious organisms, including capnophilic strains, were cultured on alternative media, according to the manufacturer's instructions, and inocula were prepared to a specified transmittance using a turbidimeter, as specified in the user guide. For each isolate, each well of a MicroPlate was inoculated with 100 µL of the cell suspension using a multichannel pipette and incubated at 37°C for 24 h. MicroPlates were read in a MicroStation semi-automated reader after 24 h and results were interpreted by the identification system's software (GEN III database, version 5.2.1). The system indicated which isolates could not be identified after 20 h and required further incubation. Reading of these isolates were performed after 3 and 6 h incubation (Wragg et al., 2014). Amplification of the 16S rRNA gene was performed using 9F and 1492R universal bacterial primers (Weisburg et al., 1991). The 16SrRNA gene sequences of related taxa were obtained from EzTaxon-e (http:// eztaxon-e.ezbiocloud.net) (Kim et al., 2012) and edited using BioEdit (Hall, 1999). Multiple alignments were performed with MUSCL (Edgar, 2004). To calculate the evolutionary distances, a two-parameter model was used (Kimura, 1983). Phylogenetic trees were constructed using the neighbor-joining method (Saitou and Nei, 1987) in MEGA5 (Tamura, 2011) with bootstrap values based on 1,000 replications (Felsenstein, 1985).

## **RESULTS AND DISCUSSION**

Based on the comparative 16S rRNA gene sequence analyses and phylogeny, nine strains, designated 15J10-8T5, KSM3-9, 15J8-3, 15J7-3T5, 15J17K, 15ML1C, 216MGT3, 216MGM6, 16MFM4 were assigned to the species level. Morphology and physiological characteristics are shown in the species description sections below.

Strains 15J10-8T5 and KSM3-9 were most closely related to *Methylobacterium hispanicum* GP34<sup>T</sup> (AJ635 304; 99.9% 16S rRNA gene sequence similarity), and *Methylobacterium soli* YIM 48816<sup>T</sup> (EU860984; 98.8%), respectively (Table 1).

Strains 15J8-3 and 15J7-3T5 were most closely related to *Microvirga subterranea* DSM 14364<sup>T</sup> (FR733708; 99.8% 16S rRNA gene sequence similarity), and *Microvirga zambiensis* WSM 3693<sup>T</sup> (HM362433; 99.2%), respectively (Table 2).

Strains 15J17K was most closely related to *Pantoea* eucrina BD  $872^{T}$  (EU216736; 99.8% 16S rRNA gene sequence similarity) (Table 3).

Strains 15ML1C, 216MGT3, 216MGM6 and 16MFM4 were most closely related to *Psychrobacter cryohalolentis* K5<sup>T</sup> (AY660685; 99.4% 16S rRNA gene sequence similarity), *Psychrobacter maritimus* Pi2-20<sup>T</sup> (AJ609272; 99.5%), *Psychrobacter okhotskensis* MD17<sup>T</sup> (AB094794; 99.7%), and *Psychrobacter urativorans* ACAM 534<sup>T</sup> (AJ609555; 99.7%), respectively (Table 4).

As expected from high 16S rRNA gene sequence similarities of the nine strains with their closest relatives, each strain formed a robust phylogenetic clade with the most closely related species (Figs. 2, 3). These results indicate that strains 15J10-8T5 and KSM3-9 are *Methylobacterium hispanicum* (Gallego *et al.*, 2005), and *Methylobacterium soli* (Cao *et al.*, 2011), repective-

**Table 2.** 16S rRNA gene sequence similarity, isolation source, medium, and incubation conditions of unrecorded strains belonging to the genus *Microvirga*

Strain ID	Most closely related species	Similarity (%)	Isolation source	Medium	Incubation conditions
15J8-3	Microvirga subterranea	99.8	Soil of Jeju	R2A	25°C, 3d
15J7-3T5	Microvirga zambiensis	99.2	Soil of Jeju	R2A	25°C, 3d

Table 3. 16S rRNA gene sequence similarity, isolation source, medium, and incubation conditions of unrecorded strain belonging to the genus *Pantoea* 

Strain ID	Most closely related species	Similarity (%)	Isolation source	Medium	Incubation conditions
15J17K	Pantoea eucrina	99.8	Soil of Jeju	R2A	25°C, 3d

Table 4. 16S rRNA gene sequence similarity, isolation source, medium, and incubation conditions of unrecorded strains belonging to the genus *Psychrobacter* 

Strain ID	Most closely related species	Similarity (%)	Isolation source	Medium	Incubation conditions
15ML1C	Psychrobacter cryohalolentis	99.4	Soil of Jeju	R2A	25°C, 3d
216MGT3	Psychrobacter maritimus	99.5	Soil of Jeju	TSA	25°C, 3d
216MGM6	Psychrobacter okhotskensis	99.7	Soil of Jeju	MA	25°C, 3d
16MFM4	Psychrobacter urativorans	99.7	Soil of Jeju	MA	25°C, 3d

TSA, Tryptic Soy Agar; MA, Marine Agar



**Fig. 2.** Neighbor-joining phylogenetic tree based on 16S rRNA gene sequences shows the relationship between the strains isolated in this study and their relatives in the genera *Methylobacterium* (1) and *Microvirga* (2). Bootstrap values (>70%) are shown above nodes for the neighbor-joining methods. Bar: 0.01 substitutions per nucleotide position.



Fig. 3. Neighbor-joining phylogenetic tree based on 16S rRNA gene sequences shows the relationship between the strains isolated in this study and their relatives in the genera *Pantoea* (1) and *Psychrobacter* (2). Bootstrap values (>70%) are shown above nodes for the neighbor-joining methods. Bar: 0.002 and 0.01 substitutions per nucleotide position, respectively.

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ly, and 15J8-3 and 15J7-3T5 are *Microvirga subterranea* (Kanso and Patel, 2011), and *Microvirga zambiensis* (Ardley *et al.*, 2012), respectively (Fig. 2). 15J17K was identified as *Pantoea eucrina* (Brady *et al.*, 2010) and 15ML1C, 216MGT3, 216MGM6 and 16MFM4 were identified as *Psychrobacter cryohalolentis* (Bakermans *et al.*, 2006), *Psychrobacter maritimus* (Romanenko *et al.*, 2004), *Psychrobacter okhotskensis* (Yumoto *et al.*, 2003) and *Psychrobacter urativorans* (Bowman *et al.*, 1996), respectively (Fig. 3).

# Description of *Methylobacterium hispanicum* 15J10-8T5

Cell is Gram-stain-negative, flagellated, and rodshaped. Colonies are pinkish-colored after 3 days of incubation on R2A at 25°C. In the GN3 microplates, acetic acid, acetoacetic acid, D-aspartic acid, bromo-succinic acid, citric acid, D-fructose, glucuronamide, L-glutamic acid, glycerol,  $\beta$ -hydroxy-D,L-butyric acid,  $\alpha$ -ketoglutaric acid, L-lactic acid, D-malic acid, L-malic acid, methyl pyruvate, mucic acid, propionic acid and D-saccharic acid are utilized as sole carbon source. But N-acetyl-D-galactosamine, N-acetyl-D-mannosamine, N-acetyl-neuraminic acid, N-acetyl-D-glucosamine, L-alanine, y-amino-butryric acid, D-arabitol, L-arginine, L-aspartic acid, D-cellobiose, dextrin, formic acid, D-fructose 6-PO4, D-fucose, L-fucose, L-galactonic acid lactone, D-galactose, D-galacturonic acid, gelatin, gentiobiose, D-gluconic acid,  $\alpha$ -D-glucose, D-glucose-6-PO4, Dglucuronic acid, L-histidine,  $\alpha$ -hydroxybutyric acid, p-hydroxy-phenylacetic acid, inosine,  $\alpha$ -keto-butyric acid, D-lactic acid methyl ester, α-D-lactose, D-maltose, D-mannitol, D-mannose, D-melibiose, \beta-methyl-D-glucoside, 3-methyl glucose, myo-inositol, pectin, glycyl-L-proline, L-pyroglutamic acid, quinic acid, D-raffinose, L-rhamnose, D-salicin, D-serine, L-serine, D-sorbitol, stachyose, sucrose, D-trehalose, D-turanose and tween 40 are not utilized.

In sensitivity tests, the tetrazolium redox dye is reduced 1% sodium lactate, aztreonam, fusidic acid, lincomycin, nalidixic acid, pH 6, rifamycin SV, sodium butyrate, tetrazolium blue and tetrazolium violet. But not in the presence of 1% NaCl, 4% NaCl, 8% NaCl, guanidine HCl, lithium chloride, minocycline, niaproof, pH 5, potassium tellurite, D-serine, sodium bromate, troleandomycin and vancomycin. Strain 15J10-8T5 (= NIBRBAC 000499690) was isolated from a soil sample, Jeju, Korea.

## Description of Methylobacterium soli KSM3-9

Cell is Gram-stain-negative, flagellated, and rodshaped. Colonies are pinkish-colored after 3 days of incubation on R2A at 25°C. In the GN3 microplates, formic acid, D-fructose 6-PO4, D-fucose, L-fucose, D-

galactose, glucuronamide, D-glucuronic acid, L-glutamic acid, glycerol, α-keto-glutaric acid, L-lactic acid, Dmalic acid, L-malic acid, methyl pyruvate, L-rhamnose, D-saccharic acid and L-serine are utilized as sole carbon source. But acetic acid, acetoacetic acid, N-acetyl-D-galactosamine, N-acetyl-D-mannosamine, N-acetyl-neuraminic acid, N-acetyl-D-glucosamine, L-alanine, y-amino-butryric acid, D-arabitol, L-arginine, D-aspartic acid, L-aspartic acid, bromo-succinic acid, D-cellobiose, citric acid, dextrin, D-fructose, L-galactonic acid lactone, Dgalacturonic acid, gelatin, gentiobiose, D-gluconic acid,  $\alpha$ -D-glucose, D-glucose-6-PO4, L-histidine,  $\alpha$ -hydroxybutyric acid, β-hydroxy-D,L-butyric acid, p-hydroxyphenylacetic acid, inosine,  $\alpha$ -keto-butyric acid, D-lactic acid methyl ester,  $\alpha$ -D-lactose, D-maltose, D-mannitol, D-mannose, D-melibiose, \beta-methyl-D-glucoside, 3methyl glucose, mucic acid, myo-inositol, pectin, propionic acid, glycyl-L-proline, L-pyroglutamic acid, quinic acid, D-raffinose, D-salicin, D-serine, D-sorbitol, stachyose, sucrose, D-trehalose, D-turanose and tween 40 are not utilized.

In sensitivity tests, the tetrazolium redox dye is reduced at 1% sodium lactate, aztreonam, fusidic acid, lincomycin, nalidixic acid, rifamycin SV, D-serine, sodium butyrate, tetrazolium blue, tetrazolium violet. But not in the presence of 1% NaCl, 4% NaCl, 8% NaCl, guanidine HCl, lithium chloride, minocycline, niaproof, pH 5, pH 6, potassium tellurite, sodium bromate, troleandomycin and vancomycin. Strain KSM3-9 (=NIBRBAC 000499691) was isolated from a soil sample, Jeju, Korea.

#### Description of Microvirga subterranea 15J8-3

Cell is Gram-stain-negative and rod-shaped. Colonies are pale yellow-colored after 3 days of incubation on R2A at 25°C. In the GN3 microplates, acetic acid, acetoacetic acid, N-acetyl-D-mannosamine, N-acetyl-D-glucosamine, D-cellobiose, dextrin, D-fructose, D-fructose 6-PO4, L-fucose, L-galactonic acid lactone, D-galactose, D-galacturonic acid, gelatin, gentiobiose, D-gluconic acid, α-D-glucose, D-glucose-6-PO4, glucuronamide, D-glucuronic acid, inosine,  $\alpha$ -keto-glutaric acid, L-lactic acid, L-malic acid, D-maltose, D-mannitol, D-mannose, 3-methyl glucose, pectin, glycyl-L-proline, L-rhamnose, L-serine, D-sorbitol, stachyose, sucrose and D-turanose are utilized as sole carbon source. But N-acetyl-D-galactosamine, N-acetyl-neuraminic acid, L-alanine, γ-aminobutryric acid, D-arabitol, L-arginine, D-aspartic acid, L-aspartic acid, bromo-succinic acid, citric acid, formic acid, D-fucose, L-glutamic acid, glycerol, L-histidine,  $\alpha$ -hydroxybutyric acid,  $\beta$ -hydroxy-D,L-butyric acid, p-hydroxy-phenylacetic acid,  $\alpha$ -keto-butyric acid, Dlactic acid methyl ester,  $\alpha$ -D-lactose, D-malic acid, Dmelibiose,  $\beta$ -methyl-D-glucoside, methyl pyruvate, mucic acid, myo-inositol, propionic acid, L-pyroglutamic acid, quinic acid, D-raffinose, D-saccharic acid, D-salicin, Dserine, D-trehalose and tween 40 are not utilized.

In sensitivity tests, the tetrazolium redox dye is reduced at 1% NaCl, 1% sodium lactate, fusidic acid, lincomycin, pH 6, potassium tellurite, sodium butyrate, tetrazolium blue, tetrazolium violet. But not in the presence of 4% NaCl, 8% NaCl, aztreonam, guanidine HCl, lithium chloride, minocycline, nalidixic acid, niaproof, pH 5, rifamycin SV, D-serine, sodium bromate, troleandomycin and vancomycin. Strain 15J8-3 (=NIBRBAC 000499692) was isolated from a soil sample, Jeju, Korea.

## Description of Microvirga zambiensis 15J7-3T5

Cell is Gram-stain-negative, flagellated, and rodshaped. Colonies are pale yellow-colored after 3 days of incubation on R2A at 25°C. In the GN3 microplates, N-acetyl-D-glucosamine, L-alanine, dextrin, D-fructose 6-PO4, D-fucose, L-fucose, L-galactonic acid lactone, D-galactose, D-galacturonic acid, glucuronamide, Dglucuronic acid, L-glutamic acid, glycerol, β-hydroxy-D,L-butyric acid, D-mannose, D-melibiose, methyl pyruvate, myo-inositol, L-rhamnose and D-sorbitol are utilized as sole carbon source. But acetic acid, acetoacetic acid, N-acetyl-D-galactosamine, N-acetyl-D-mannosamine, N-acetyl-neuraminic acid, y-amino-butryric acid, D-arabitol, L-arginine, D-aspartic acid, L-aspartic acid, bromo-succinic acid, D-cellobiose, citric acid, formic acid, D-fructose, gelatin, gentiobiose, D-gluconic acid, α-D-glucose, D-glucose-6-PO4, L-histidine, α-hydroxybutyric acid, p-hydroxy-phenylacetic acid, inosine, α-keto-butyric acid, α-keto-glutaric acid, L-lactic acid, D-lactic acid methyl ester,  $\alpha$ -D-lactose, D-malic acid, L-malic acid, D-maltose, D-mannitol, β-methyl-D-glucoside, 3-methyl glucose, mucic acid, pectin, propionic acid, glycyl-L-proline, L-pyroglutamic acid, quinic acid, D-raffinose, D-saccharic acid, D-salicin, D-serine, Lserine, stachyose, sucrose, D-trehalose, D-turanose and tween 40 are not utilized.

In sensitivity tests, the tetrazolium redox dye is reduced at 1% NaCl, aztreonam, fusidic acid, lincomycin, nalidixic acid, pH 6, rifamycin SV, sodium butyrate, tetrazolium blue and tetrazolium violet. But not in the presence of 1% sodium lactate, 4% NaCl, 8% NaCl, guanidine HCl, lithium chloride, minocycline, niaproof, pH 5, potassium tellurite, D-serine, sodium bromate, troleandomycin and vancomycin. Strain 15J7-3T5 (= NIBR BAC000499693) was isolated from a soil sample, Jeju, Korea.

#### Description of Pantoea eucrina 15J17K

Cell is Gram-stain-negative, flagellated, and rodshaped. Colonies are pale lemon-colored after 3 days

of incubation on R2A at 25°C. In the GN3 microplates, acetic acid, acetoacetic acid, N-acetyl-D-mannosamine, N-acetyl-D-glucosamine, L-alanine, γ-amino-butryric acid, L-arginine, L-aspartic acid, bromo-succinic acid, D-cellobiose, citric acid, dextrin, formic acid, D-fructose, D-fructose 6-PO4, D-fucose, L-fucose, L-galactonic acid lactone, D-galactose, D-galacturonic acid, gentiobiose, D-gluconic acid, α-D-glucose, D-glucose-6-PO4, glucuronamide, D-glucuronic acid, L-glutamic acid, glycerol, L-histidine,  $\alpha$ -hydroxybutyric acid, inosine,  $\alpha$ -ketobutyric acid, L-lactic acid, D-lactic acid methyl ester,  $\alpha$ -D-lactose, D-malic acid, L-malic acid, D-maltose, D-mannitol, D-mannose, D-melibiose, β-methyl-D-glucoside, 3-methyl glucose, mucic acid, myo-inositol, propionic acid, glycyl-L-proline, quinic acid, L-rhamnose, D-saccharic acid, D-salicin, D-serine, L-serine, D-sorbitol, sucrose and D-trehalose are utilized as sole carbon source. But N-acetyl-D-galactosamine, N-acetyl-neuraminic acid, D-arabitol, D-aspartic acid, gelatin,  $\beta$ -hydroxy-D,L-butyric acid, p-hydroxy-phenylacetic acid, α-keto-glutaric acid, methyl pyruvate, pectin, L-pyroglutamic acid, D-raffinose, stachyose, D-turanose and tween 40 are not utilized.

In sensitivity tests, the tetrazolium redox dye is reduced at % NaCl, 1% sodium lactate, 4% NaCl, 8% NaCl, guanidine HCl, lincomycin, lithium chloride, niaproof, pH 5, pH 6, potassium tellurite, rifamycin SV, sodium bromate, tetrazolium blue, tetrazolium violet and troleandomycin. But not in the presence of aztreonam, fusidic acid, minocycline, nalidixic acid, D-serine, sodium butyrate and vancomycin. Strain 15J17K (=NIBRBAC 000499695) was isolated from a soil sample, Jeju, Korea.

## Description of Psychrobacter cryohalolentis 15ML1C

Cell is Gram-stain-negative and coccus-shaped. Colonies are pale yellow-colored after 3 days of incubation on R2A at 25°C. In the GN3 microplates, acetic acid, N-acetyl-neuraminic acid, L-alanine, y-amino-butryric acid, D-arabitol, L-aspartic acid, D-cellobiose, dextrin, D-fructose, D-fructose 6-PO4, D-fucose, L-fucose, D-galacturonic acid, gelatin,  $\alpha$ -D-glucose, D-glucose-6-PO4, glucuronamide, L-glutamic acid, L-histidine, β-hydroxy-D,L-butyric acid, L-lactic acid, D-lactic acid methyl ester,  $\alpha$ -D-lactose, D-malic acid, D-maltose, D-mannitol, D-mannose, D-melibiose, β-methyl-D-glucoside, 3-methyl glucose, myo-inositol, pectin, propionic acid, glycyl-L-proline, D-raffinose, L-rhamnose, L-serine, D-sorbitol, stachyose and tween 40 are utilized as sole carbon source. But acetoacetic acid, N-acetyl-D-galactosamine, N-acetyl-D-mannosamine, N-acetyl-D-glucosamine, L-arginine, D-aspartic acid, bromo-succinic acid, citric acid, formic acid, L-galactonic acid lactone, D-galactose, gentiobiose, D-gluconic acid, D-glucuronic

acid, glycerol,  $\alpha$ -hydroxybutyric acid, p-hydroxy-phenylacetic acid, inosine,  $\alpha$ -keto-butyric acid,  $\alpha$ -keto-glutaric acid, L-malic acid, methyl pyruvate, mucic acid, L-pyroglutamic acid, quinic acid, D-saccharic acid, D-salicin, D-serine, sucrose, D-trehalose and D-turanose are not utilized.

In sensitivity tests, the tetrazolium redox dye is reduced at 1% NaCl, 1% sodium lactate, 4% NaCl, 8% NaCl, aztreonam, lincomycin, lithium chloride, nalidixic acid, pH 6, potassium tellurite, rifamycin SV, D-serine, sodium bromate, sodium butyrate, tetrazolium blue and troleandomycin. But not in the presence of fusidic acid, guanidine HCl, minocycline, niaproof, pH 5, tetrazolium violet and vancomycin. Strain 15ML1C (=NIBRBAC 000499696) was isolated from a soil sample, Jeju, Korea.

#### **Description of Psychrobacter maritimus 216MGT3**

Cell is Gram-stain-negative and rod-shaped. Colonies are pale yellow-colored after 3 days of incubation on Tryptic Soy Agar at 25°C. In the GN3 microplates, acetic acid, acetoacetic acid, N-acetyl-D-glucosamine, Lalanine, γ-amino-butryric acid, L-aspartic acid, bromosuccinic acid, citric acid, dextrin, formic acid, D-fructose, D-fructose 6-PO4, D-fucose, L-fucose, L-galactonic acid lactone, D-galacturonic acid, gelatin, gentiobiose, D-gluconic acid, α-D-glucose, glucuronamide, D-glucuronic acid, L-glutamic acid, L-histidine, α-hydroxybutyric acid, β-hydroxy-D,L-butyric acid, p-hydroxy-phenylacetic acid,  $\alpha$ -keto-butyric acid,  $\alpha$ -keto-glutaric acid, L-lactic acid, D-lactic acid methyl ester,  $\alpha$ -D-lactose, D-malic acid, L-malic acid, D-maltose, D-mannitol, D-mannose, D-melibiose, β-methyl-D-glucoside, mucic acid, pectin, propionic acid, glycyl-L-proline, L-pyroglutamic acid, L-rhamnose, D-serine, L-serine, D-sorbitol, stachyose, sucrose, D-trehalose, D-turanose and tween 40 are utilized as sole carbon source. But N-acetyl-D-galactosamine, N-acetyl-D-mannosamine, N-acetylneuraminic acid, D-arabitol, L-arginine, D-aspartic acid, D-cellobiose, D-galactose, D-glucose-6-PO4, glycerol, inosine, 3-methyl glucose, methyl pyruvate, myo-inositol, quinic acid, D-raffinose, D-saccharic acid and Dsalicin are not utilized.

In sensitivity tests, the tetrazolium redox dye is reduced at 1% NaCl, 1% sodium lactate, 4% NaCl, 8% NaCl, aztreonam, lincomycin, lithium chloride, pH 6, potassium tellurite, rifamycin SV, sodium butyrate and tetrazolium blue. But not in the presence of fusidic acid, guanidine HCl, minocycline, nalidixic acid, niaproof, pH 5, D-serine, sodium bromate, tetrazolium violet, troleandomycin and vancomycin. Strain 216MGT3 (= NIBR BAC000499697) was isolated from a soil sample, Jeju, Korea.

#### Description of Psychrobacter okhotskensis 216MGM6

Cell is Gram-stain-negative, flagellated, and rodshaped. Colonies are pale yellow-colored after 3 days of incubation on Marine Agar at 25°C. In the GN3 microplates, acetic acid, acetoacetic acid, N-acetyl-D-glucosamine, L-alanine, y-amino-butryric acid, D-arabitol, L-arginine, D-aspartic acid, L-aspartic acid, D-cellobiose, dextrin, D-fructose, D-fructose 6-PO4, D-fucose, Lfucose, L-galactonic acid lactone, D-galactose, D-galacturonic acid, gentiobiose, D-gluconic acid,  $\alpha$ -D-glucose, D-glucose-6-PO4, glucuronamide, D-glucuronic acid, L-glutamic acid, glycerol, L-histidine,  $\alpha$ -hydroxybutyric acid,  $\beta$ -hydroxy-D,L-butyric acid,  $\alpha$ -keto-glutaric acid, L-lactic acid, D-lactic acid methyl ester, D-malic acid, L-malic acid, D-maltose, D-mannitol, D-mannose, Dmelibiose,  $\beta$ -methyl-D-glucoside, 3-methyl glucose, methyl pyruvate, mucic acid, propionic acid, glycyl-Lproline, L-pyroglutamic acid, quinic acid, L-rhamnose, D-saccharic acid, D-salicin, L-serine, D-sorbitol, sucrose, D-trehalose, D-turanose and tween 40 are utilized as sole carbon source. But N-acetyl-D-galactosamine, N-acetyl-D-mannosamine, N-acetyl-neuraminic acid, bromo-succinic acid, citric acid, formic acid, gelatin, p-hydroxy-phenylacetic acid, inosine,  $\alpha$ -keto-butyric acid,  $\alpha$ -D-lactose, myo-inositol, pectin, D-raffinose, D-serine and stachyose are not utilized.

In sensitivity tests, the tetrazolium redox dye is reduced at 1% NaCl, 1% sodium lactate, 4% NaCl, 8% NaCl, lincomycin, lithium chloride, pH 6, potassium tellurite, rifamycin SV, sodium butyrate, tetrazolium blue and tetrazolium violet. But not in the presence of aztreonam, fusidic acid, guanidine HCl, minocycline, nalidixic acid, niaproof, pH 5, D-serine, sodium bromate, troleandomycin and vancomycin. Strain 216MGM6 (= NIBR BAC000499698) was isolated from a soil sample, Jeju, Korea.

## Description of Psychrobacter urativorans 16MFM4

Cell is Gram-stain-negative and coccus-shaped. Colonies are pale yellow-colored after 3 days of incubation on Marine Agar at 25°C. In the GN3 microplates, acetic acid, acetoacetic acid, *N*-acetyl-D-galactosamine, L-alanine,  $\gamma$ -amino-butryric acid, D-arabitol, L-arginine, D-aspartic acid, L-aspartic acid, bromo-succinic acid, citric acid, dextrin, formic acid, D-fructose, D-fructose 6-PO4, D-fucose, L-fucose, L-galactonic acid lactone, D-galactose, D-galacturonic acid, gelatin, gentiobiose, D-gluconic acid,  $\alpha$ -D-glucose, D-glucose-6-PO4, glucuronamide, D-glucuronic acid, L-glutamic acid, L-histidine,  $\alpha$ -hydroxybutyric acid,  $\beta$ -hydroxy-D,L-butyric acid,  $\alpha$ -keto-butyric acid,  $\alpha$ -bractose, D-malic acid, L-malic acid, D-maltose, D-mannose, D-melibiose, β-methyl-D-glucoside, 3-methyl glucose, methyl pyruvate, mucic acid, pectin, propionic acid, glycyl-L-proline, L-pyroglutamic acid, quinic acid, D-raffinose, L-rhamnose, D-saccharic acid, D-serine, L-serine, stachyose and tween 40 are utilized as sole carbon source. But *N*-acetyl-D-mannosamine, *N*-acetyl-neuraminic acid, *N*-acetyl-D-glucosamine, D-cellobiose, glycerol, p-hydroxy-phenylacetic acid, inosine, D-mannitol, myoinositol, D-salicin, D-sorbitol, sucrose, D-trehalose and D-turanose are not utilized.

In sensitivity tests, the tetrazolium redox dye is reduced at 1% NaCl, 1% sodium lactate, 4% NaCl, 8% NaCl, aztreonam, lincomycin, lithium chloride, pH 6, potassium tellurite, rifamycin SV, D-serine, sodium butyrate and tetrazolium blue. But not in the presence of fusidic acid, guanidine HCl, minocycline, nalidixic acid, niaproof, pH 5, sodium bromate, tetrazolium violet, troleandomycin and vancomycin. Strain 16MFM4 (= NIBR BAC000499699) was isolated from a soil sample, Jeju, Korea.

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# REFERENCES

- Ardley, J.K., M.A. Parker, S.E. De Meyer, R.D. Trengove, G.W. O'Hara, W.G. Reeve, R.J. Yates, M.J. Dilworth, A. Willems and J.G. Howieson. 2012. *Microvirga lupini* sp. nov., *Microvirga lotononidis* sp. nov. and *Microvirga zambiensis* sp. nov. are alphaproteobacterial root-nodule bacteria that specifically nodulate and fix nitrogen with geographically and taxonomically separate legume hosts. Int. J. Syst. Evol. Microbiol. 62:2579-2588.
- Bakermans, C., H.L. Ayala-del-Río, M.A. Ponder, T. Vishnivetskaya, D. Gilichinsky, M.F. Thomashow and J.M. Tiedje. 2006. *Psychrobacter cryohalolentis* sp. nov. and *Psychrobacter arcticus* sp. nov., isolated from Siberian permafrost. Int. J. Syst. Evol. Microbiol. 56:1285-1291.
- Bowman, J.P., J. Cavanagh, J.J. Austin and K. Sanderson. 1996. Novel Psychrobacter species from Antarctic ornithogenic soils. Int. J. Syst. Evol. Microbiol. 46:841-848.
- Brady, C.L., I. Cleenwerck, S.N. Venter, K. Engelbeen, P. De Vos and T.A. Coutinho. 2010. Emended description of the genus *Pantoea*, description of four species from human clinical samples, *Pantoea septica* sp. nov., *Pantoea eucrina* sp. nov., *Pantoea brenneri* sp. nov. and *Pantoea*

*conspicua* sp. nov., and transfer of *Pectobacterium cypripedii* (Hori 1911) Brenner *et al.* 1973 emend. Hauben *et al.* 1998 to the genus as *Pantoea cypripedii* comb. nov. Int. J. Syst. Evol. Microbiol. 60:2430-2440.

- Cao, Y.-R., Q. Wang, R.X. Jin, S.K. Tang, Y. Jiang, W.X. He, H.X. Lai, L.H. Xu and C.L. Jiang. 2011. *Methylobacterium soli* sp. nov. a methanol-utilizing bacterium isolated from the forest soil. Antonie Leeuwenhoek 99:629-634.
- Doetsch, R. 1981. Determinative methods of light microscopy. Manual of Methods for General Bacteriology: 21-33.
- Edgar, R.C. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput. Nucleic Acids Res. 32:1792-1797.
- Felsenstein, J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. Evolution 39:783-791.
- Gallego, V., M.T. García and A. Ventosa. 2005. Methylobacterium hispanicum sp. nov. and Methylobacterium aquaticum sp. nov., isolated from drinking water. Int. J. Syst. Evol. Microbiol. 55:281-287.
- Hall, T.A. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/ NT. In: Nucleic acids symposium series, 1999. vol 41. [London]: Information Retrieval Ltd., c1979-c2000., pp 95-98.
- Kanso, S. and B.K. Patel. 2003. *Microvirga subterranea* gen. nov., sp. nov., a moderate thermophile from a deep subsurface Australian thermal aquifer. Int. J. Syst. Evol. Microbiol. 53:401-406.
- Kim, O.-S., Y.J. Cho, K. Lee, S.H. Yoon, M. Kim, H. Na, S.C. Park, Y.S. Jeon, J.H. Lee, H. Yi, S. Won and J. Chun. 2012. Introducing EzTaxon-e: a prokaryotic 16S rRNA gene sequence database with phylotypes that represent uncultured species. Int. J. Syst. Evol. Microbiol. 62:716-721.
- Kimura, M. 1983. The neutral theory of molecular evolution. Cambridge University Press.
- Romanenko, L.A., A.M. Lysenko, M. Rohde, V.V. Mikhailov and E. Stackebrandt. 2004. *Psychrobacter maritimus* sp. nov. and *Psychrobacter arenosus* sp. nov., isolated from coastal sea ice and sediments of the Sea of Japan. Int. J. Syst. Evol. Microbiol. 54:1741-1745.
- Saitou, N. and M. Nei. 1987. The neighbor-joining method: a new method for reconstructing phylogenetic trees. Mol. Biol. Evol. 4:406-425.
- Tamura, K., D. Peterson, N. Peterson, G. Stecher, M. Nei and S. Kumar. 2011. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods Mol. Biol. Evol. 28:2731-2739.
- Weisburg, W.G., S.M. Barns, D.A. Pelletier and D.J. Lane. 1991. 16S ribosomal DNA amplification for phylogenetic study. J. Bacteriol. 173:697-703.
- Williams, K.P., J.J. Gillespie, B.W. Sobral, E.K. Nordberg, E.E. Snyder, J.M. Shallom and A.W. Dickerman. 2010. Phylogeny of gammaproteobacteria. J. Bacteriol. 192:

2305-2314.

- Williams, K.P., B.W. Sobral and A.W. Dickerman. 2007. A robust species tree for the alphaproteobacteria. J. Bacteriol. 189:4578-4586.
- Woese, C.R. 1987. Bacterial evolution. Microbiological reviews 51:221-271.
- Wragg, P., L. Randall and A. Whatmore. 2014. Comparison of Biolog GEN III MicroStation semi-automated bacterial identification system with matrix-assisted laser desorption ionization-time of flight mass spectrometry and 16S ribosomal RNA gene sequencing for the iden-

tification of bacteria of veterinary interest. J. Microbiol. Methods 105:16-21.

Yumoto, I., K. Hirota, Y. Sogabe, Y. Nodasaka, Y. Yokota and T. Hoshino. 2003. *Psychrobacter okhotskensis* sp. nov., a lipase-producing facultative psychrophile isolated from the coast of the Okhotsk Sea. Int. J. Syst. Evol. Microbiol. 53:1985-1989.

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