# One new species and one new record of lymantriine moths (Lepidoptera: Erebidae: Lymantriinae) in Korea

Nang-Hee Kim<sup>1</sup>, Sei-Woong Choi<sup>1,\*</sup> and Sung-Soo Kim<sup>2</sup>

<sup>1</sup>Department of Environmental Education, Mokpo National University, Muan, Jeonnam 58554, Republic of Korea

Herein, we report a new record of *Arna bipunctapex* (Hampson) and a new species, *Euproctis fulvatus* sp. nov. in Korea; both are Lymantriine moths. *Arna bipunctapex* is distinguished by a relatively large wingspan with two black dots and a small dot between these black dots in the apical region of a yellowish forewing. The male genitalia of *A. bipunctapex* can be distinguished by the lack of a process on the sacculus of valva while the female genitalia can be distinguished by an antrum that is basally flat with two lateral digitate arms. *Euproctis fulvatus* is distinguished by yellowish wings with a medially curved central fascia of the forewing. The male genitalia can be distinguished by the bifid, digitate uncus and the simple, square-shaped valva with a distal strong invaginated margin. The female genitalia can be distinguished by the long, medially twisted, ductus dursae with simple antrum posteriorly strongly sclerotized and ovate corpus bursae without signum. Larvae of *E. fulvatus* are distinguished by a black head with a pair of long, black, lateral tufts, dorsum with 10–11 white intersegmental dots and bright red setal warts on T2–A8, and bright red glands on A6 and A7.

Keywords: Erebidae, Korea, Lepidoptera, Lymantriinae, taxonomy

© 2019 National Institute of Biological Resources DOI:10.12651/JSR.2019.8.3.288

## Introduction

The subfamily Lymantriinae comprises over 2,900 species in 360 genera and occurs primarily in the Old World tropics (Kitching and Rawlins, 1998). Synapomorphies of the subfamily are recognized: the prespiracular counter-tympanal hood and the strongly reduced proboscis in adults and the dorsal glands at the center of the sixth and seventh abdominal segments in larvae (Zahiri *et al.*, 2010). Recently Kim *et al.*(2016) published a checklist of the Noctuoidea from Korea, listing 50 species in 25 genera of Lymantriinae moths. In addition to that list, herein, we report two species of Lymantriinae for the first time in Korea: *Arna bipunctapex* (Hampson) and *Euproctis fulvatus* sp. nov.

## MATERIALS AND METHODS

Adult moths were collected at night after entering a UV-light bucket trap (BioQuip, USA). All collected adults were preserved in a freezer and then mounted for examination. Larvae were collected through direct observation

of plants and were reared at the laboratory until eclosion. For slide preparation of male and female genitalia, each specimen was prepared by boiling the abdomen in 10% KOH for approximately 20 min. Subsequently, scales and tissues were removed, the remaining sclerotized structure was stained with chlorazol black, and mounted on slides in Euparal solution. Wingspan measurement was the distance from the tip of the left forewing to the tip of the right forewing.

Genomic DNA was extracted from moth legs using the DNeasy Blood and Tissue Extraction Kit (Qiagen, UK), according to the manufacturer's instructions. The obtained gDNA, the 658-bp 5' end region of the COI gene was amplified, using a polymerase chain reaction (PCR) method. For amplification, we used a previously described standard barcoding primer pair: LCO1490, 5'-GGT CAA CAA ATC ATA AAG ATA TTG G-3' and HCO2198, 5'-TAA ACT TCA GGG TGA CCA AAA AAT CA-3' (Folmer *et al.*, 1994). The PCR was conducted using FastMix French PCR kits (i-Taq; iNtRon Biotechnology, Korea) under the following conditions: initial denaturation for 3 min at 94°C, followed by 30 cycles of 94°C for 1 min, 50°C for 30 sec, and 72°C for 1 min, with a subse-

<sup>&</sup>lt;sup>2</sup>Research Institute for East Asian Environment and Biology, Gangdong-gu, Seoul 05264, Republic of Korea

<sup>\*</sup>Correspondent: choisw@mokpo.ac.kr

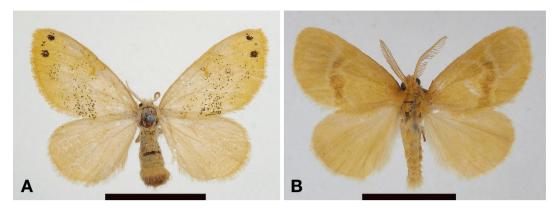


Fig. 1. Adults of two new Lymantriine species from Korea. Scale bar = 10 mm. A. Arna bipunctapex (Hampson), B. Euproctis fulvatus sp. nov.

quent final 7 min extension at 72°C. Electrophoresis was carried out using 1 × TAE buffer on 1% agarose gel with Top Green Nucleic Acid Gel Stain (LED; Genomic Base, Korea) for 15 min at 135 V to confirm successful DNA amplification. The obtained PCR products were purified with a PCR purification kit (iNtRON) and were sequenced with forward and reverse primers (Genotech Korea). Sequences were edited manually to check ambiguous bases by applying forward and reverse primer sequences using the Seqman program (DNASTAR Lasergene software, version 7.1; DNASTAR, USA). The DNA barcoding sequence was compared with publicly available sequence databases, such as Biotechnology Information (NCBI) and GenBank, and through BLAST (http://blast.ncbi.nlm. nih.gov/Blast.cgi) and the Barcode of Life Data (BOLD) system (http://boldsystems.org; Ratnasingham and Hebert, 2007).

Terminology related to adults and larvae, including male and female genitalia refers to Lafontaine and Fibiger (2006) and Wagner (2005). All materials have been deposited in the Insect Collection, Department of Environmental Education, Mokpo National University, Korea. Abbreviations: GW, Gangwon-do; GB, Gyungsangbuk-do; GN, Gyungsangnam-do; JB, Jeollabuk-do; and JN, Jeollanam-do.

## Systematic Accounts

Order Lepidoptera Linnaeus, 1758 Family Erebidae Leach, [1815] Subfamily Lymantriinae Hampson, 1893 Genus *Arna* Walker, 1855

#### Arna bipunctapex (Hampson)

두점노랑독나방(신청) (Figs. 1A, 2A)

Somena bipucntapex Hampson, 1891: 57. TL: [Inida]

Nilgiri.

Artaxa bipunctapex: Kirby, 1892: 453. Aroa bipunctapex: Swinboe, 1892: 191. Euproctis bipunctapex: Collenette, 1932: 60. Nygmia bipunctapex: Swinhoe, 1922: 483. Arna bipunctapex: Holloway, 1999: 46.

**Material examined.** 1♀, Chopyeong-gil, Gunoe-myeon, JN: Wando, N34°21′38.30″ E126°39′51.93″, 77 m, 5.XI.2015. SS Kim.

**Diagnosis.** Distinguished by the relatively large wingspan (27 mm) with two black dots in the apical region of the forewing. Head with pectinate female antennae with short pectinations, broad and yellowish frons and porrect and long (about twice the eye diameter) labial palpi. Forewing ground color light yellow-brown, tinged with brown scales on the central fascia and a postmedian, and apical region with two black dots. Hindwing ground color light yellow tinged with brown scales. Female genitalia (Fig. 2A). Papillae anales broad with rounded outer margin; anterior apophyses almost equal length to posterior apophyses; antrum U-shaped, basally flat, with digitate arms laterally; ductus dursae long, twice the corpus bursae; and corpus bursae ovate without signum.

**Distribution.** Korea, Japan, China, Thailand, India, Nepal, Malaysia, Singapore.

Remarks. Worldwide, 12 species of *Arna* have been described. *Arna* larvae feed on *Sepium*, *Terminalis*, *Shorea*, and *Caerya* (Holloway, 1999). Up to now, two species of *Arna* are known in Korea: *A. pseudoconspersa* (Strand, 1914) and *A. bipunctapex*. These two species are similar in the shapes of their central fascia and the two black apical dots on the forewing. *A. bipunctapex* can be distinguished from *A. pseudoconspersa* by the presence of a small dot between two blackish dots in the apical region of the forewing. The male genitalia of *A. bipunctapex* can be distinguished from those of *A. pseudoconspersa* by the absence of a process on the sacculus of valva in *A. bipunctapex* (Wang *et al.*, 2010; 2011). The female geni-

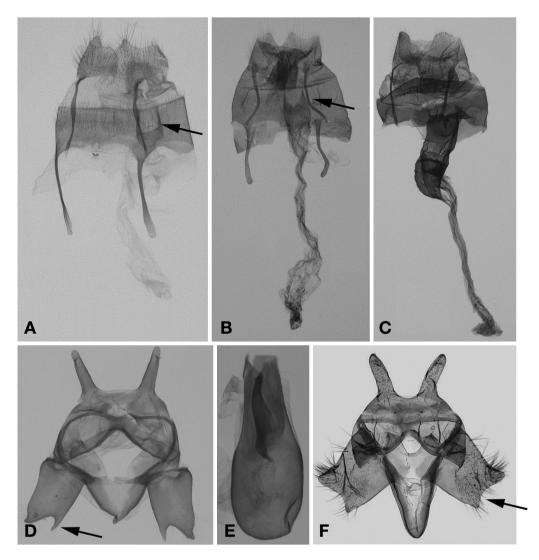


Fig. 2. Male and female genitalia of four Lymantriine species from Korea. A-C. Female genitalia, D-F. Male genitalia. A. *Arna bipunctapex* (Hampson), B. *Arna pseudoconspersa* (Strand), C-E. *Euproctis fulvatus* Choi and Kim sp. n. F. *Euproctis wilemani* Collenette (from Wang *et al.*, 2010).

talia of *A. bipunctapex* can be distinguished from those of *A. pseudoconspersa* (Fig. 2B) by the shape of the antrum, basally flat with two lateral digitate arms in *A. bipunctapex*.

Genus Euproctis Haworth, 1809

*Euproctis fulvatus* Kim, Choi and Kim sp. nov 띠무늬독나방(신칭) (Figs. 1B, 2C-E, 3)

**Type.** Holotype. male, Ssangyoung-ri, Hanbando-myeon, GW: Yeongwol, 3.IX.2005. SS Kim. Paratypes. 1♂, Chuncheon-si, GW: Chuncheon, 3.VIII.1998. SS Kim; 1♂ Huindol-ro, Byeonggok-myeon, GB: Yeongdeok, N36°36′24″, E129°24′49″ 14 m, 3.VI.2015. SS Kim;

1♂, Daedae-ri, Yueo-myon, GN: Changnyeong, N35° 33′07″, E128°24′53″ 5 m, 29.VIII.2009. SS Kim; 1♂, Daedae-ri, Yueo-myon, GN: Changnyeong, N35°33′07″, E128°24′53″ 5 m, 29.VIII.2009. SS Kim; 1♂, Sinseong-ri, Seongnae-myeon, JB: Gochang, N35°32′55″, E126° 43′10″ 12 m, 17.VI.2016. SS Kim; 1♀, Guseong-ri, Sani-myeon, JN: Haenam, N34°41′27.43″ E126°24′29.96″, 12 m, 25.VIII.2016. SW Choi; 2♂1♀, Guseong-ri, Sani-myeon, JN: Haenam, N34°41′27.43″ E126°24′29.96″, 3 m, ex. Larvae 20.IV.2017, ex pupa 8−9.VI.2017. NH Kim. **Diagnosis.** Distinguished by the yellowish wing color with a medially curved central forewing fascia. Dimorphic size (wingspan male 19−25 mm, female 33 mm). Head covered with yellow scales; antenna bipectinate with long pectinations in male and short pectinations in



Fig. 3. Last instar of Euproctis fulvatus sp. nov.

female; frons broad and yellowish; labial palpi porrect, short and approximately equal to the eye diameter. Forewing dark yellowish; central fascia ochreous, slender, bar-shaped, medially curved. Hindwing yellowish white and termen dark yellowish. Abdomen dark yellow. Male genitalia (Fig. 2D, E): Uncus bifid, digitate, apically with a short triangular process; saccus V-shaped; valva simple, square, distally strongly invaginated; aedeagus stout, anterior broad; one long sclerotized cornutus present. Female genitalia (Fig. 2C) Papillae anales broad; ductus dursae long, medially twisted, posteriorly strongly sclerotized with simple antrum; corpus bursae ovate without signum. Larvae (Fig. 3). Distinguished by a black head with a pair of long, black, lateral tufts, dorsum with 10-11 white intersegmental dots and bright red setal warts on T2-A8, and bright red glands on A6 and A7. Larva to 3

**Biology.** This species feeds on *Vicia angustifolia* var. *segetilis* (L.) Schreb. (1835) (Fabales). Three larvae on *Vicia* were collected on April 20 and 21, 2018, molted twice (April 28 and May 6), pupated May 19 and eclosed June 8 and 9. The species is bivoltine, flying in June and August–September in Korea.

## Distribution. Korea.

**Remarks.** This new species is similar to *Euproctis wile-mani* Collenette, but can be distinguished by the shape of male genitalia (Fig. 2F). The DNA sequence of *E. fulvatus* (Appendix 1) is similar to that of *E. pyraustis* Meyrick (GenBank accession number: KP081919), but the nucleotide sequence of *E. pyraustis* has a 9.6% sequence divergence from *E. fulvatus*.

#### **ACKNOWLEDGEMENTS**

We thank Dr. Alexander Schintlmeister for the information on the Lymantriine moths of China. This study was supported by a grant from the National Institute of

Biological Resources (NIBR, (NIBR 201801201), funded by the Ministry of Environment (MOE) of the Republic of Korea.

### REFERENCES

- Collenette, C.L. 1932. The Lymantriidae of the Malay Peninsula. Novitates Zoologicae 38:49-102.
- Folmer, O., M. Black, W. Hoeh, R. Lutz and R. Vrijenhoek. 1994. DNA primers for amplification of mitochondrial cytochrome *c* oxidase subunit I from diverse metazoan invertebrates. Molecular Marine Biology and Biotechnology 3:294-299.
- Hampson, G.F. 1891. Illustrations of typical specimens of Lepidoptera Heterocera in the collection of the British Museum. Part VIII- The Lepidoptera Heterocera of the Nilgiri District. London. P. 57, pl. 140, f. 13.
- Holloway, J.D. 1999. The Moths of Borneo [Part 5]: Family Lymantriidae. Malayan Nature Journal 53:1-188.
- Kim, S.S., S.W. Choi, V. Kononenko, A. Schintlmeister and J.C. Sohn. 2016. Revised list of the Korean Noctuoidea based on latest classification. Entomological Research Bulletin 32:138-160.
- Kirby, W.F. 1892. A synonymic catalogue of Lepidoptera Heterocera (moths). Sphinges and Bombyces. Gurney & Jackson, London Volume 1. pp. 1-951.
- Kitching, I.J. and J.E. Rawlins. 1998. The Noctuoidea. In: N.P. Kristensen (ed.), Handbook of Zoology/Handbuch der Zoologie, Lepidoptera, Moths and Butterflies, Vol. 1. Evolution, Systematics and Biogeography. W. de Gruyter, Berlin. pp. 355-401.
- Lafontaine, J.D. and M. Fibiger. 2006. Revised higher classification of the Noctuoidae (Lepidoptera). The Canadian Entomologist 138:610-635.
- Ratnasingham, S. and P.D.N. Hebert. 2007. BOLD: the barcode of life data system (www.barcodinglife.org). Molecular Ecology Notes 7:355-364.
- Swinhoe, C. 1892. Catalogue of eastern and Australian Lepidoptera Heterocera in the collection of the Oxford University Museum. Part 1 Sphinges and Bombyces. Claredon Press, Oxford, 324 pp.
- Swinhoe, C. 1922. A revision of the genera of the family Liparidae. Annals and Magazine of Natural History 10(9):449-484.
- Wagner, D.L. 2005. Caterpillars of Eastern North America. Princeton University Press, Princeton.
- Wang, H.S., X.L. Fan and M. Wang. 2010. A New Record of *Euproctis wilemani* (Lepidoptera: Lymantriidae) from Hainan Island. Florida Entomologist 93(2):325-326.
- Wang, H.S., M. Wang and X.L. Fan. 2011. Notes on the tribe Nygmiini (Lepidoptera: Erebidae: Lymantriinae) from Nanling National Nature Reserve, with description of a new species. Zootaxa 2887:57-68.
- Zahiri, R., I.J. Kitching, J.D. Lafontaine, M. Mutanen, L.

Kaila, J.D. Holloway and N. Wahlberg. 2010. A new molecular phylogeny offers hope for a stable family level classification of the Noctuoidea (Lepidoptera). Zoologica Scripta 40:158-173.

Submitted: December 27, 2018 Revised: July 1, 2019 Accepted: July 8, 2019

**Appendix 1.** DNA sequence of *Euproctis fulvatus* sp. nov. Sequence alignment of DNA barcoding region from *Euproctis fulvatus* sp. nov and *E. pyraustis*. The alignment was generated by the Clustal W method. Different sequences are presented.

			30			60
Euproctis fulvatus	AACACTTTAT	TTTATTTTTG	GAATTTGAGC	AGGAATAGTC	GGCTCATCTT	TAAGAATAAT
Euproctis pyraustis				T	G	
			90			120
Euproctis fulvatus	AATTCGAACT	GAATTAGGAA	ACCCGGGATC	TTTTATCAAT	AACGATCAAA	TCTTTAATAC
Euproctis pyraustis	TC		CTT	T	T	.T
			150			180
Euproctis fulvatus	TATTATTACT	GCTCATGCTT	TTATTATAAT	TTTTTTTATA	GTTATACCTA	TTATAATCGG
Euproctis pyraustis		A			A.	T
			210			240
Euproctis fulvatus	AGGGTTTGGA	AATTGATTAG	TACCATTAAT	ATTAGGAGCT	CCTGATATAG	CTTTCCCCCG
Euproctis pyraustis	A		.T	C		.ATT
			270			300
Euproctis fulvatus	AATAAATAAT	ATAAGATTTT	GACTTTTACC	CCCCTCCTTA	ATTCTTCTTT	TATCAAGAAG
Euproctis pyraustis	Т			ATT	C	.CT
			330			360
Euproctis fulvatus	AATTATTGAA	AATGGAGCTG	GAACTGGATG	AACTGTTTAT	CCTCCTCTTT	CTTCTAATAT
Euproctis pyraustis	• • • • • • • • • •	A.	.TC		C	
			390			420
Euproctis fulvatus	TGCCCATAGA	GGAAGATCAG	TCGACTTAAC	TATTTTTCT	TTACATCTTG	CTGGAATTTC
Euproctis pyraustis	G		.TT	c	С.Т	
			450			480
Euproctis fulvatus	TTCAATTTTA	GGGGCAATTA	ATTTTATTAC	CACTATTATC	AATATACGTT	TAAATAATTT
Euproctis pyraustis	A	A		AT		
	3 mg3 mmmg3 m	C111 m1 CCmm	510	1 CC1 CMMCC1	3 mm3 G3 GGGm	540
Euproctis fulvatus	ATCATTTGAT	CAAATACCTT	TATTTGTTTG	AGCAGTTGGA	ATTACAGCCT	TCCTACTTCT
Euproctis pyraustis	T			<b>T</b>	T.	.TT 600
Formula Colombia	TTTATCTCTC	CCCCM3 MMCC	CCGGAGCTAT	TACTATGTTA	TTAACTGATC	GAAATTTAAA
Euproctis fulvatus		CCCGTATTGG		AA		GAAATTTAAA
Euproctis pyraustis		TA.	.т 630	AA		658
Funnantia fuluation	TACTTCATTT	TTTGATCCTG	CAGGAGGGG	AGATCCAATT	TTATATCAAC	658 TTTATTTA
Euproctis fulvatus Euproctis pyraustis		CA.	A			.C
Euprocus pyraustis		C A.	A			